

Vegetable *milks* and their fermented derivative products

NEUS BERNAT^{a*}, MAITE CHÁFER^a, AMPARO CHIRALT^a, AND CHELO GONZÁLEZ-MARTÍNEZ^a

^a Instituto de Ingeniería de Alimentos para el Desarrollo. Universitat Politècnica de València, Camino de Vera s/n, 46022 Valencia, Spain

*Corresponding author
neuberpe@upvnet.upv.es

Received: 16 July 2013; Published online: 18 April 2014

Abstract

The so-called vegetable *milks* are in the spotlight thanks to their lactose-free, animal protein-free and cholesterol-free features which fit well with the current demand for healthy food products. Nevertheless, and with the exception of soya, little information is available about these types of *milks* and their derivatives. The aims of this review, therefore, are to: highlight the main nutritional benefits of the nut and cereal vegetable *milks* available on the market, fermented or not; describe the basic processing steps involved in their manufacturing process; and analyze the major problems affecting their overall quality, together with the current feasible solutions. On the basis of the information gathered, vegetable *milks* and their derivatives have excellent nutritional properties which provide them a high potential and positive market expectation. Nevertheless, optimal processing conditions for each raw material or the application of new technologies have to be researched in order to improve the quality of the products. Hence, further studies need to be developed to ensure the physical stability of the products throughout their whole shelf-life. These studies would also allow for a reduction in the amount of additives (hydrocolloids and/or emulsifiers) and thus reduce the cost of the products. In the particular case of fermented products, the use of starters which are able to both improve the quality (by synthesizing enhanced flavors and providing optimal textures) and exert health benefits for consumers (i.e. probiotics) is the main challenge to be faced in future studies.

Keywords: Nut *milk*; Cereal *milk*; Processing; Fermentation

Abbreviations

CLSM	Confocal laser scanning microscopy
CVD	Cardiovascular diseases
DM-2	Diabetes mellitus type 2;
GI	Glycemic index
MCFA	Medium-chain fatty acids
MUFA	Mono-unsaturated fatty acids
PUFA	Poly-unsaturated fatty acids
SFA	Saturated fatty acids
UHPH	Ultra high pressure homogenization
UHT	Ultra high temperature

1 Introduction

Nowadays, there is a global awareness of nutrition-related chronic diseases. In its 2009 annual report on global health risks, the World Health Organization (WHO) determined the distribution of deaths attributable to 19 leading risk factors worldwide. More than half of these factors were nutrition-related: blood pressure due to sodium consumption, cholesterol, obesity, deficiencies of iron and zinc, among others (Stuckler & Basu, 2011). Increasingly, consumers are more aware of the relationship between nutrition and health. Indeed, newly designed foods are not only intended to satisfy hunger and provide nutrients for humans, but also to prevent nutrition-related chronic diseases and to improve well-being, both physical and mental (Burdock & Carabin, 2008; Granato, Branco, Nazzaro, Cruz, & Faria, 2010; S. Kaur & Das, 2011; Ozen, Pons, & Tur, 2012). This trend is justified if several factors are considered, such as an increase in public health awareness (a consequence of a more highly educated population), an aging population and their desire for improving the quality of their later years, an increase in healthcare costs, advances in research and technology or changes in government regulations and accountability. The food market reflects to an ever greater degree the consumer demand for healthy food products. A clear example of this tendency can be seen in the so-called vegetable *milks*, which are mainly made of nuts and cereals and have a long history in both Eastern and Western cultures. European sales of soya *milk* and other non-dairy *milks* are increasing by over 20% per year; Spain being the EU country in which the non-dairy

drinks market grew the most (Organic Monitor, 2006). Similarly, total USA retail sales of soy, almond, rice and other plant milks reached \$1.3 billion in 2011 (Packaged Facts, 2012).

The best known and most popular vegetable *milk* derives from soy, although the demand for almond, rice, oat and coconut *milks* is on the increase. Wide ranges of nut and cereal vegetable *milks* are currently available on the market in a broad array of formulations: flavored, sweetened/unsweetened, low-fat and/or fortified. Excluding Asia, non-dairy *milk* alternatives (vegetable *milks*) still represent a relatively small market overall; nonetheless, the growing awareness of allergy and intolerance issues and the lactose-free, cholesterol-free and low-calorie positioning of these products are bringing about a rise in purchase levels (Stone, 2011). In fact, marketing strategies of those products focus on comparing their health benefits with those of dairy products. Furthermore, experts are starting to consider possible relations between vegetable products and the prevention of cancer, atherosclerosis or inflammatory diseases, since free radicals play a key role in those pathologies and these types of food are an excellent source of antioxidants (Scalbert & Williamson, 2000). The lactose intolerant and/or those people allergic to cow milk are prime consumers of these types of *milks*, but they are also in great demand by people without health problems, such as vegans and vegetarians.

The development and further increase in demand of such products would have an extra advantage, which could be of economic interest for many countries: the raw material they de-

rive from (nuts and cereals) do not generally require specific soil nor climatic conditions, they are able to adapt to different climates although, of course, the productivity might change (Osca, 2007; Coniglio, 2008). For example, almond tree farming is considered to be a dry cultivation with low soil fertility, low rainfall and minimum pruning and plant protection requirements (Navarro-Muñoz, 1996; Saura, Cañellas, & Soler, 1988). Oat is a temperate crop which grows well in damp, marginal upland areas (Welch & McConnell, 2001). These facts would benefit the rapid implementation of these raw materials in non-cultivated lands around the world and maybe, this could contribute to the rural development of developing countries and allow these vegetable products to attain highly competitive prices within the world market.

Taking into account the positive trends of these products in the food market and bearing in mind that the literature contains little information about them, the aims of this work are to: highlight the main nutritional benefits of these kinds of *milks*, fermented or not; describe the basic processing steps involved in their manufacturing process; and analyze the major problems affecting the overall quality, together with the possible solutions currently available. Therefore, this review focuses on the study of nut and cereal vegetable *milks* available on the market and their fermented derivatives.

2 Types of nut and cereal vegetable *milks* and their nutritional benefits

All the commercial vegetable *milks* share common features such as being lactose-free, animal protein-free or cholesterol-free. Taking into account the raw materials and their nutritional and health properties, vegetable *milks* can be broadly classified in two large differentiated groups: nut and cereal *milks*. Both kinds of products are in the state of the art owing to the new-knowledge impact of their compounds on some current chronic diseases, such as cardiovascular diseases (CVD), type 2 Diabetes mellitus (DM-2), obesity and some cancers. These metabolic diseases are linked with our daily lifestyle, notably an unbal-

anced energy-rich diet, lacking in fiber and protective bioactive compounds, such as micronutrients and phytochemicals (Fardet, 2010). All these limited nutrients commented on above are readily available in both cereals and nuts.

Apart from nuts and cereals, other raw materials have been used industrially, such as tubers (e.g. tigernuts) and plants (e.g. hemp, sunflower...). However, these milky based products are only well accepted in specific countries. Despite its local commerce, tigernut *milk* has also been explained in detail in this review due to its interesting composition and health properties.

2.1 Cereal grains and their *milks*

Cereals are known as a good source of the necessary daily energy, vitamins, several minerals, dietary fiber and phytochemicals, including phenolic compounds, carotenoids, vitamin E, lignans, inulin, starch, sterols and phytates (Okarter & Liu, 2010; J. L. Ward et al., 2008). The chemical composition of those cereals whose vegetable *milk* has been commercialized is summarized in Table 1. With respect to the supply of vitamins, cereals are considered an important source of group B vitamins, especially thiamin, riboflavin, folates and niacin (McKevith, 2004). Dietary fiber is present in large quantities and this is rich in fructo-oligosaccharides, which are reportedly effective at stimulating the growth of *Bifidobacteria* and *Lactobacilli* in the human intestine (K. D. Kaur, Jha, Sabikhi, & Singh, 2011). Besides this prebiotic effect, their phenolic compounds have also been reported to possess gastroprotective properties, in addition to their antioxidant, cholesterol-lowering, anti-atherogenic, anti-carcinogenic and anti-inflammatory effects (C. Chen et al., 2004; Dykes & Rooney, 2006; Prior & Gu, 2005). Indeed, epidemiological studies have shown an association between increased wholegrain consumption and reduced risks of various types of chronic diseases, such as CVD, obesity, DM-2 and some cancers (Chan, Wang, & Holly, 2007; de Munter, Hu, Spiegelman, Franz, & van Dam, 2007; Esmailzadeh, Mirmiran, & Azizi, 2005; Larsson, Giovannucci, Bergkvist, & Wolk, 2005; Mellen, Walsh, & Herrington, 2008; Murtaugh, Jacobs, Jacob, Steffen, & Marquart,

2003; Schatzkin, Park, Leitzmann, Hollenbeck, & Cross, 2008; van de Vijver, van den Bosch, van den Brandt, & Goldbohm, 2009). More specific properties and health benefits of each cereal grain whose vegetable *milk* has been commercialized are summarized in Table 4.

In order to gain the greatest benefit from the health properties of cereals, several aspects have to be considered. For instance, it is important to use and consume whole grain and not the refined, since most of the health components are located in the bran and germ. So, the use of the whole grain is highly recommended when producing the cereal *milks*. Another point to consider is the anti-nutrient content in some cereals, primarily phytic acid (mineral chelator) or saponins (toxic in high amounts and bitter tasting), although their presence can be reduced and/or eliminated by pre-treatment processes such as grinding, soaking, heat treatments, fermentations and germinations (Brady, Ho, Rosen, Sang, & Karwe, 2007; Sharma & Kapoor, 1996; Zhu et al., 2002). Despite the anti-nutritional components, so beneficial are whole-grain cereal's properties that important food authorities, such as the U.S. Department of Agriculture (USDA), have strongly recommended 6-11 servings of grain products daily (Dewanto, Wu, & Liu, 2002).

2.2 Nuts and nut milks

Due to their composition, nuts and nut-based products have recently attracted a great deal of attention from food, nutrition and health specialists. Table 1 shows the chemical composition of those nuts whose vegetable *milks* have been commercially produced. Nuts are rich in mono- (MUFA) and polyunsaturated fatty acids (PUFA), vegetable proteins, dietary fiber, phytosterols, polyphenols, vitamins and minerals (Phillips, Ruggio, & Ashraf-Khorassani, 2005; Segura, Javierre, Lizarraga, & Ros, 2006). Most of those compounds have antioxidant properties and are proven to provide a beneficial effect on plasma lipid profile, low-density lipoprotein (LDL) oxidation and inflammatory processes, among others (Carlson, Eisenmann, Norman, Ortiz, & Young, 2011; Egert, Kratz, Kannen-

berg, Fobker, & Wahrburg, 2011; Gillingham, Harris-Janz, & Jones, 2011; Jones et al., 2011; Liu, 2012; Myers & Allen, 2012; H. A. Ward et al., 2012; Whent et al., 2012). Additionally, Vinson and Cai (2012) analyzed the antioxidant efficacy in different nuts, obtaining the following order of importance: walnut > cashew > hazelnut \approx almond. Epidemiological studies have linked frequent nut consumption to a reduced risk of CVC, DM-2 or death by all-cause mortality (Kelly & Sabate, 2006). Moreover, Li et al. (2009) observed that an increase in nut consumption was significantly associated with a more favorable plasma lipid profile, including lower LDL cholesterol, total cholesterol and apolipoprotein B-100 concentrations; but they did not observe significant associations with non-high-density lipoprotein (HDL) cholesterol or inflammatory markers. Furthermore, nuts have a high K/Na ratio, which contributes to maintain well-balanced electrolytes in the human body, and, in addition to the prebiotic effect of their dietary fiber commented on above, the carbohydrates from nuts are complex (low Glycemic Index (GI)), which help to maintain blood sugar at healthy levels.

In spite of the fact that around 50% of a nut is made up of lipids, regular nut consumption within a balanced diet has been shown to improve humans' lipid profile, increase endothelial function and reduce inflammation, without causing weight gain (C.-Y. Chen, Lapsley, & Blumberg, 2006; R. D. Mattes, Kris-Etherton, & Foster, 2008; Salas-Salvado, Casas-Agustench, Murphy, Lopez-Uriarte, & Bullo, 2008; Zambón et al., 2000). Thus, in addition to providing both nutrients and bioactive antioxidants, nut milks may be a useful dietary tool for reducing risk factors that cause diseases with a major mortality rate in developed countries, such as metabolic syndrome, DM-2 or CVD. Indeed, the U.S. Food and Drug Administration approved a health claim between nuts and heart disease, suggesting that 42 g per day of most nuts as part of a low saturated-fat and cholesterol diet may reduce the risk of heart disease (FDA. U.S. Food and Drug Administration, 2003). The European Food Safety Authority (EFSA) also published a scientific opinion on the substantiation of health claims related to nuts and essential fatty acids (omega-

Table 1: Composition of raw materials commercially used for producing vegetable *milks*. Average values shown are expressed per 100 g of edible part. Nutrient Recommended Daily Allowances corresponding to a healthy adult with standard physical activity and lifestyle are also included

Name	NUTS			CEREALS										RDA ^X			
	Almond ^A	Chestnut ^A	Hazelnut ^B	Walnut ^B	Amaranth ^B	Barley ^B	Corn ^B	Kamut ^B	Millet ^B	Oat ^A	Quinoa ^A	Rice ^A (brown)	Sesame ^D		Spelt ^B	Coconut ^B	Tigernut ^A
Energy KJ (KCal)	2453 -589	939 -225	2629 -628	2738 -654	1554 -371	1346 -352	1528 -365	1411 -337	1583 -378	1670 -401	1276 -306	1607 -386	2660 -644	1415 -338	1482 -354	1706 -409	8374 -2000
Lipids (g)	54.65 35.01	5.3 0.6	60.75 45.65	65.21 8.93	7.02 1.685	1.16 0.15	4.74 1.25	2.2 0.214	4.22 0.773	6.9 2.2	5.56 1.4	2.64 0.83	49.7 18.8	2.43 0.445	33.49 1.425	23.74 16.47	<70
MUFPA (g)	12.28	1.3	7.92	4.717	2.778	0.56	2.16	0.616	2.134	2.5	2.1	0.89	21.8	1.258	0.366	2.21	<20
SFA (g)	4.93	3.2	4.46	6.13	1.459	0.25	0.67	0.192	0.723	1.2	0.5	0.52	7.6	0.406	29.698	4.02	<20
Cholesterol(mg)	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	50
Protein (g)	19.13	4	14.95	15.23	13.56	9.91	9.42	14.7	11.02	16.9	13.8	7.5	17.7	14.57	3.33	6.13	50
moisture (g)	5.87	4.48	5.31	4.07	11.29	10.09	10.37	10.95	8.67	0	11.5	11.4	4.24	11.02	46.99	n.a.	270
Carbohydrates(g)	6.2	39.7	16.70	13.71	65.25	7.7	74.26	70.38	72.85	66.3	49.2	81.3	9.28	70.19	15.23	42.54	
sugars (g)	5.3	7.9	4.34	2.61	1.69	0.8	0.64	n.a.	n.a.	n.a.	5.92	0.45	0.45	n.a.	6.23	n.a.	
starch (g)	0.11	31.8	n.a.	2.1	57.27	73.3	73.3	n.a.	79.7	43.27	7.9	7.9	7.9	10.7	9	17.4	25
Dietary fiber (g)	8.35	7.1	9.7	6.7	6.7	15.6	7.3	9.1	8.5	10.6	7.9	3	0	0	0	0	800
Vitamin A** (µg)	0	0	1	1	0	0	11	1	0	0	0	0	n.a.	n.a.	n.a.	10	12
Vitamin E** (µg)	24	1.4	15.03	0.7	1.19	0.2	0.49	0.6	0.05	0.7	0.45	0.6	n.a.	0.79	0.24	6	80
Vitamin C (mg)	n.a.	0	6.3	1.3	4.2	0	0	0.591	0.421	0.76	0	0	0.791	0	3.3	0.23	1.1
Thiamin (mg)	0.21	0.18	0.643	0.34	0.116	0.19	0.385	0.178	0.29	0.14	0.4	0.08	0.247	0.113	0.066	0.1	1.4
Riboflavin (mg)	0.78	0.1	0.113	0.15	0.2	0.114	0.20	0.364	4.72	0.96	n.a.	6.8	4.52	6.843	0.54	1.8	50
Niacin (mg)	5.3	n.a.	1.125	1.125	0.923	4.61	3.64	6.35	4.72	0.96	n.a.	0.51	0.79	0.23	0.054	n.a.	200
Vitamin B6 (mg)	0.11	0.3	0.563	0.537	0.591	0.26	0.62	0.255	0.384	n.a.	0.2	0.51	0.79	0.23	0.054	n.a.	2.5
Biotin (µg)	64	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	30	40	97	45	26	141	200
Folate (µg)	70	85	113	98	82	23	19	n.a.	85	0	0	0	0	0	0	0	2.5
Vitamin B12 (µg)	0.44	n.a.	n.a.	0	n.a.	0	0	n.a.	0	1.35	0	0	0.05	n.a.	n.a.	n.a.	6
Pantothenic acid (mg)	10.36	10.6	11	2	4	9	35	6	5	2	61	1.5	2.31	8	20	37.63	<6
Sodium (mg)	767.25	591.6	680	441	508	280	287	446	195	429	780	223	468	388	356	519.2	2000
Potassium (mg)	248.25	47.3	114	98	159	29	210	24	8	54	79	21	96	27	14	69.54	800
Calcium (mg)	524.88	87.6	290	346	557	221	210	386	285	523	230	303	604	401	113	232.23	700
phosphorous (mg)	258.13	163	163	158	248	79	127	134	114	177	210	143	324	136	32	86.88	375
Magnesium (mg)	3.59	2.91	4.7	2.91	7.61	2.5	2.71	4.41	3.01	4.72	7.8	1.7	14.6	4.44	2.43	3.41	14
Iron (mg)	3.6	0.6	2.45	3.09	2.87	2.13	2.21	3.68	1.68	3.97	3.3	1.8	5.74	3.28	1.1	4.19	10
Zinc (mg)	1	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	0.63	n.a.	0.85	1.58	n.a.	n.a.	n.a.	1
copper (mg)	1	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	2	<51.9	n.a.	n.a.	n.a.	55
Selenium (µg)	4	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	6	n.a.	22	<5	n.a.	n.a.	n.a.	150
Iodide (µg)	2	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	1.23	n.a.	n.a.	n.a.	2
Manganese (µg)	2.54	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	1.23	n.a.	n.a.	n.a.	2

A: Data extracted from Spanish Food Composition Database (RedBDDCA)
 B: Data extracted from United States Department of Agriculture Database (USDA)
 C: Data extracted from French agency for food environmental and occupational health and safety Database (Anses)
 X: Recommended Daily Allowances Data extracted from European Directive 90/496/CEE
 RDA: Recommended Daily Allowances
 *: Retinol equivalents
 **: alpha-tocopherol equivalents
 n.a.: data not available
 MUFPA: mono-unsaturated fatty acids
 PUFA: poly-unsaturated fatty acids
 SFA: saturated fatty acids

3/omega-6) in nut oil, which is related to anti-inflammatory, heart health, weight management and healthy cardiovascular system effects (Agostoni et al., 2011).

Although coconut is commonly classified as a nut, its composition does not follow the trend of this food group (Table 1), which means that not all of the above-mentioned health properties of nuts are associated with it. As can be seen in the chemical composition of coconut (Tables 1 and 4), the particularity of this traditional *milk* from the Asian, African and Pacific regions is its medium chain fatty acid (MCFA) lipid profile, which is similar to human milk (Chiewchan, Phungamngoen, & Siriwattayothin, 2006); the most predominant is lauric acid (45-53% of total coconut fats) and this MCFA was reported to be antibacterial, antiviral and antifungal (Raghavendra & Raghavarao, 2010). In spite of the lipid profile being mostly saturated, Enig (2004) reported that MCFA are absorbed directly from the intestine and sent straight to the liver to be rapidly metabolized for energy production and, thus, they do not participate in the biosynthesis and transport of cholesterol. Furthermore, the high amount of antioxidants determines the long shelf-life of this vegetable *milk* and is good for the health. Other interesting health benefits are summarized in Table 4.

Tigernuts are another interesting source of raw material to be used for the production of vegetable *milks*. The major components (Table 1) of this tuber are complex carbohydrates, mainly starch and dietary fiber, which provide vegetable *milk* with low GI. Furthermore, the protein content is rich in arginine, which liberates hormones that produce insulin; thus being suitable for diabetics (Adejuyitan, 2011). Besides its antioxidant compounds, the lipid profile of tigernuts is similar to that found in olive oil; therefore, the derived *milk* has a positive effect on the cholesterol level. Other interesting health benefits are detailed in Table 4.

More specific properties and health benefits of each nut whose derived vegetable milk has been commercialized are summarized in Table 4.

3 Vegetable milk processing

Industrial vegetable *milk* processing is based on five main steps: grinding, water extraction, filtration, homogenization and pathogen removal treatment. Nevertheless, depending on the raw material and the desired final product characteristics, the process slightly differs. Thus, the processing is subsequently explained separately, taking into account the different groups of these above-mentioned *milks* commented upon.

3.1 Cereal *milk* processing

A typical flow diagram of cereal *milk* processing is shown in Figure 1. Before going through the extraction procedure, cereals are conditioned: this mostly refers to husking, washing and grain classification. Pre-conditioning is a requisite for Quinoa grain due to its saponins' content (toxic in high amounts and imparts a bitter taste). Mechanical abrasion and/or washing are sufficient to remove this unwanted compound (Brady et al., 2007).

Once the cereal grain is conditioned, it is submitted to a coarse dry grinding to facilitate the subsequent water extraction. This water extraction process is usually carried out in colloidal mills by adding hot water and the ground cereal at the same time. The colloidal mills are used to reduce the particle size of the solid in aqueous suspension by applying high levels of hydraulic shear to the process liquid. The amount of water added would depend on the final pre-established concentration. This step is carried out in hot conditions, on one hand, to ease the extraction of soluble compounds and, on the other hand, to provoke starch gelatinization and improve the subsequent enzymatic treatment. The main difference observed between these and other types of vegetable *milks* is this enzymatic treatment which they are submitted to after the extraction procedure. This step is needed to attain the low viscosity which the consumer demands of these products. This viscosity is provided mainly by starch and other polysaccharides after the thermal treatments.

After removing the non-extracted solids, the milky liquid obtained is then temperature ad-

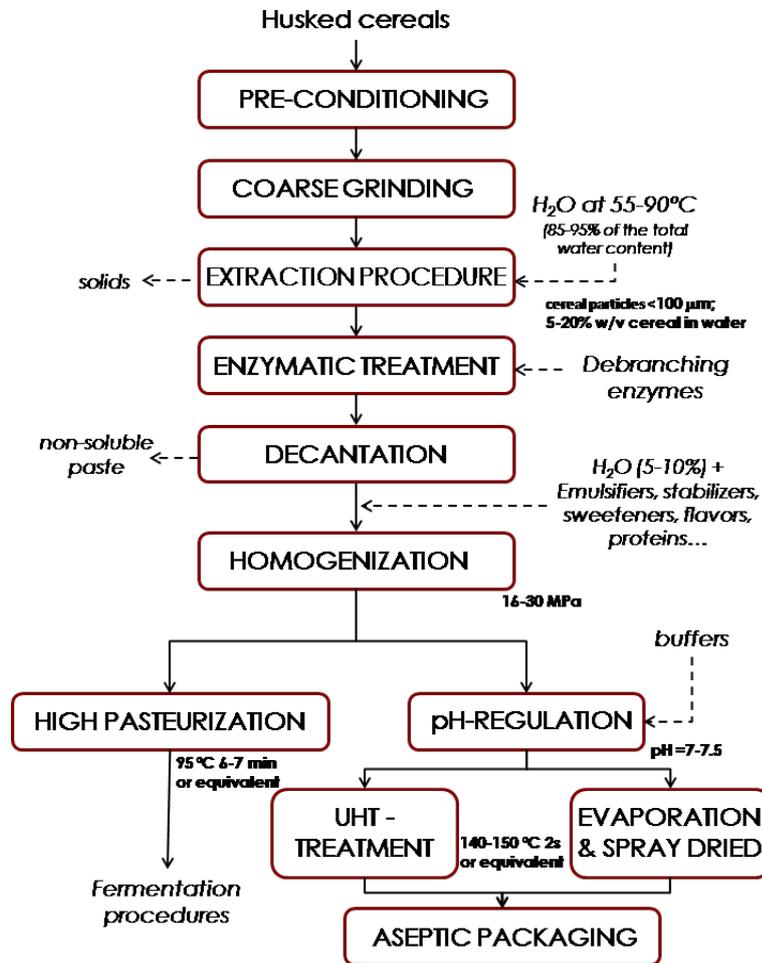


Figure 1: Typical cereal *milk* processing flow diagram.

justed and pH adjusted, with appropriate buffers, to the optimum level for the enzymatic treatment. The pH values, temperature and the time-remaining would depend on the types of enzymes used and those would be chosen taking into account the final product's desired viscosity, sugar content and/or texture parameters. Generally, enzyme composition comprises mainly α -amylase and β -amylase activities and is devoid of β -glucanase and proteinase activities, since glucans and proteins have interesting nutrient value and their degradation is not desirable (Triantafyllou, 2002). When the desired textural properties are reached, a homogenization process

is applied to ensure physical stabilization during the product's shelf life. The usual homogenization pressures range between 16-30 MPa, although some researchers are studying the use of ultra-high pressures (UHPH, >150 MPa) in vegetable *milk* production (Bernat, Chafer, A., & González-Martínez, 2011; N. Cruz et al., 2007; Valencia-Flores, Hernandez-Herrero, Guamis, & Ferragut, 2013). Food additives, such as emulsifiers (lecithin), stabilizers (hydrocolloids), sweeteners, either natural (sucrose, fructose or glucose syrups from agave, corn, rice or wheat) or synthetic (acesulfame K, aspartame or sucralose) and, sometimes, flavoring agents (cocoa, soluble

coffee, vanilla or cinnamon) are often introduced before the homogenization step. The amounts of these additives incorporated ranged from 0.4-2.5 % (w/v) in emulsifiers, 0.025-0.3 % (w/v) in stabilizers, 5-8 % (w/v) in sweeteners and 0.5-3 % (w/v) in flavoring agents (Erra, 2012; Pereyra & Mutilangi, 2012; Triantafyllou, 2002; Marti, Martinez, Miralles, & Perez, 2010). Further information on the ingredients used to formulate already commercialized cereal milks is shown in Table 2.

To ensure quality and safety, after having readjusted the pH to standard values of *milks* with buffers such as sodium carbonates, potassium carbonates, sodium hydroxides or potassium diphosphates (Erra, 2012), homogenized *milks* are either heat treated or spray dried and finally aseptically packaged. Heat parameters of temperature, time and pressure would be stipulated taking into account the type of product, the particle size, viscosity, initial microbial load and stability of components under thermal conditions. The inactivation conditions of the enzymes previously used is also a variable when choosing heat parameters, since this step is also used to eradicate the activities of residual enzymes. Ultra High Temperature (UHT) treatment is commonly chosen (140-150 °C, 2 sec) or high pasteurization (95°C > 6 min or equivalent treatment) might also be used when the *milks* product is to be fermented (Erra, 2012; Pereyra & Mutilangi, 2012; Marti et al., 2010; Triantafyllou, 2002).

The final milky product chemical composition of major components (protein, lipids and carbohydrates) in different types of cereals is summarized in Table 2. Considering these *milks* are to be used as substitutes for cow milk, it is remarkable that there is a high fiber content and a low lipid content with a better profile than standard milk.

3.2 Nut milk processing

The general industrial flow diagram of nut *milk* production is presented in Figure 2. The steps shown are mostly the same as in cereal *milk* processing but the enzymatic treatment is removed, since the low starch content of these nuts do not confer a negative effect on the viscosity of the final product.

Nut conditioning consists of washing and selection, plus a blanching treatment in order to facilitate both the further peeling of nuts and the initial microbial load reduction. In view of the fact that no enzymes are used, possible food additives such as emulsifiers, sweeteners, hydrocolloids and/or flavors can be introduced during the grinding step, and thereby facilitate the optimal dispersion of ingredients. The types of food additives chosen are the same as the ones mentioned in cereal *milk* processing.

The final milky product chemical composition of major components (proteins, lipids and carbohydrates) in different types of tree nuts is summarized in Table 2. In spite of the fact that the over 50 % of a nut's content is made up of lipids (Table 1), final contents end up with lower values than non-defatted cow milk, which varies from 3 to 5 % (w/v). Moreover, considering these vegetable *milks* are to be used for the same purposes as cow milk, the (MUFA+PUFA)/SFA ratio (SFA, Saturated Fatty Acids) is much higher than other animal milks and, hence, they are healthier. Almond *milk* stands out among other nut *milks* as being an appropriate alternative to cow milk, since, besides the lipid profile, it has a low ratio of Na/K and a balanced ratio of Ca/P (Luengo, 2009).

In the production of tigernut *milk*, the pre-conditioning step is more complex: in the extraction procedure, tigernuts need to be softened prior to the milling process (by rehydration with water for around 18 hours) and a preliminary germicidal treatment (active chlorine) is required to decrease the initial microbial load (it is a tuber) (CRDO: Consejo Regulador de la Denominación de Origen Chufa de Valencia, 2012; Sanful et al., 2009). Also, an enzymatic treatment is required due to the high starch content of this nut, as has been commented on above for the cereal *milks* manufacture.

In industrial coconut *milk* production, the coconut meal has to be obtained by shelling, paring and washing. After that, coconut meal is submitted to a blanching process reinforced with chemical agents, such as NaHSO₃, for different purposes: facilitate the removal of the brown testa, enhance oxidation stability due to inner enzyme denaturation and facilitate further grinding. Once coconut meat is ground and water

Table 2: Chemical composition of commercial vegetable milks. Average values shown are expressed per 100 mL of liquid product

VEGETABLE MILK	INGREDIENTS	BASIC CHEMICAL COMPOSITION (g 100 mL ⁻¹)										MAIN MANUFACTURERS
		Proteins	TOTAL	MUFAs	PUFA	SEFA	TOTAL	Carbohydrates	SIGARS	Dietary fibre	Sodium	
Almond	Water, almonds (7-8%), thickener (corn starch/maltodextrin), stabilizer (carrageenan/gellan gum), emulsifier (sunflower/soya lecithin), salt, almond flavor	1.1-1.8	1.46-2.6	0.8-1	0.42-1.4	0.3-0.6	1-8	0.1-1.6	0.4-4.2	0.01-0.06	(1), (2), (6), (7), (10), (12), (13), (14), (15)	
		0.05	3.5	2.3	0.8	0.5	6.2	0.3	2.4	0.03	-1	
Chestnut	Water, chestnut (14%), sweetener (rice syrup/ agave syrup), thickener (maltodextrin), almond oil, natural flavor, salt	0.6-0.8	1.5-2.8	2	0.1	0.7	6.5-8	3.2-6	0.42-0.9	0.175	(1), (15)	
		1.2	1	0.3	0.6	0.1	7.4	1.7	0.4	0.02	(1), (12)	
Hazelnut Walnut	Water, hazelnut (7%), agave syrup, thickener (corn maltodextrin)	0.6	1.9	1.1	0.3	0.5	8	n.a.	0.3	0.05	-1	
		0.5	1	0.3	0.6	0.1	11	7	0.5	0.09	-3	
Amaranth	Water, amaranth (17%), sunflower oil, salt	0.52	1.4	0.7	0.5	0.2	9.5	6.5	0.6	0.03	(3), (4), (6), (12)	
		0.65-0.7	0.8-1.45	0.2-0.4	0.5-0.88	0.1-0.21	7.5-20	4.6-7.1	0.4-0.5	0.04-0.09	(2), (3), (4), (5), (6), (7), (8), (9), (10), (11), (12), (15), (16)	
Barley	Water, barley (17%), sunflower oil, salt	0.7	1.2	0.3	0.8	0.1	9.8	5.7	0.4	0.09	(9), (10), (11), (12), (15), (16)	
		0.5-1	0.7-1.6	0.8+0.099	0.2-0.4	0.1-0.3	4.75-11.8	1.5-5.5	0.44-0.9	0.04-0.1	(1), (2), (6)	
Corn	Water, white corn (17%), sunflower oil, salt	1.5	2.8	1.5	0.6	0.7	3.7	3.4	0.6	0.2	(2), (3), (4), (5), (6), (7), (8), (11), (12), (15), (16)	
		0.12-0.8	0.9-3	0.38-1	0.11-1.7	0.1-0.37	4.6-15.6	4.3-7.1	0.12-0.15	0.04-0.1	-1	
Kamut	Water, rice (12-20%), stabilizer (carrageenan/xanthan gum), sunflower/soya lecithin	0.6	2.4	1.4	0.5	0.5	6.7	n.a.	0.2	0.01	(2), (3), (5), (6)	
		0.2-0.7	1-1.4	0.3-0.37	0.8-0.87	0.1-0.16	8.4-9.6	4.7	0.2-0.7	0.05-0.13	(2), (3), (5), (6)	
Millet	Water, millet (12-14%), sunflower oil, salt	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	
Oat	Water, oat (8-14%), sunflower oil, salt	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	
Quinoa	Water, quinoa (7-14%), sweetener (agave syrup), thickener (corn maltodextrin), stabilizer (pectin/carrageenan), sunflower oil, emulsifier (soya lecithin)	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	
Rice	Water, rice (12-20%), stabilizer (carrageenan/xanthan gum), sunflower/soya lecithin, salt	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	
Sesame	Water, sesame (7%), sweetener (agave syrup), thickener (corn maltodextrin), high oleic sunflower oil	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	
Spelt	Water, spelt (14-15%), sunflower oil, salt	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	
Coconut	Coconut cream(10%), sweetener (cane sugar), natural flavor, stabilizer (carrageenan/guar gum), salt	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	
Tigernut	Water, tigernuts (10-14%), sweetener (agave syrup/sucralose), thickener (corn maltodextrin/lupinosa starch), stabilizer (carrageenan/ carboxymethyl cellulose)	0.2-0.7	1.9-2.4	0.1	n.a.	1.4-2.3	0.9-5.7	0.4-1	0.4-1.1	0.02-0.1	(13), (14)	
		0.4-0.7	2-2.4	1.1-1.6	0.19-0.3	0.4-0.6	5.3-8.3	4.5-6.5	0.57-1	0.02-0.06	(1), (3), (12)	

n.a.: data not available
 PUFA: polyunsaturated fatty acids
 MUFA: mono-unsaturated fatty acids
 SEFA: saturated fatty acids

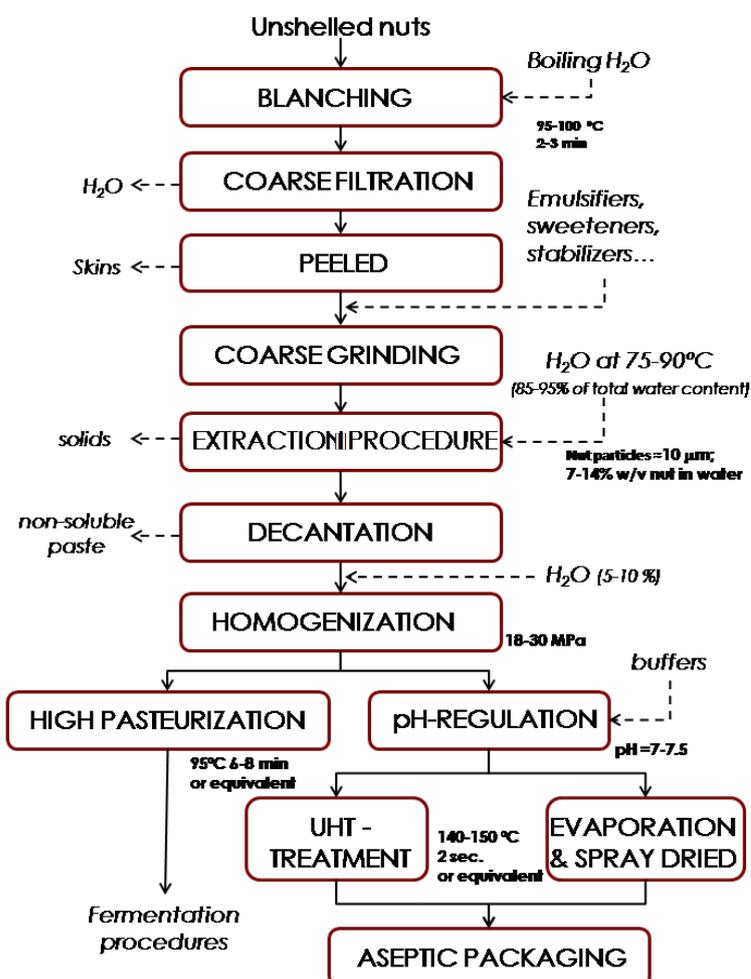


Figure 2: Typical nut *milk* processing flow diagram.

extraction is developed, the milky liquid is finally filtrated using double layers of cheese cloth (Mepba, Achinewhul, & Pillay, 2009).

The major technological problem found during the processing or shelf life of these cereals and nut derived *milks* is related with the low physical stability of the liquid dispersion, usually with low viscosity, which promotes the phase separation of the unstable fat globules caused by flocculation and coagulation phenomena in a short period of time. Moreover, fibers and non-soluble material will also separate, either by sedimentation or floatation, thus contributing to the watery effect of the product. The employment of

an optimal thermal treatment and homogenization pressures during the *milk* processing, the addition of both amphiphilic compounds and hydrocolloids or the use of UHPH could contribute to the development of an excellent product with desirable sensory attributes. Using UHPH allows for a longer shelf life of the product, since greater physical stability is achieved mainly due to a reduction in the size of the fat globule that prevents coalescence. Figure 3 shows pictures of hazelnut *milk* obtained by means of the Confocal laser scanning microscopy (CLSM) technique in which the effect of UHPH is proved; as can be seen, almost all the fat globules in untreated

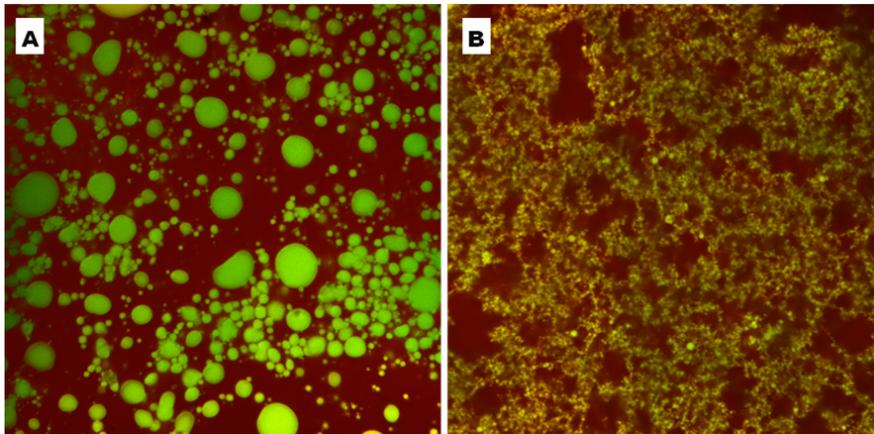


Figure 3: Confocal pictures of hazelnut *milk* non-treated (A) and homogenized at 172 MPa (B), where fat globules appeared green-yellow colored. *Source*: personal compilation, University of Valencia

milk (Fig. 3A) are aggregated, while in that which has been UHPH-treated (Fig 3A), not only are the fat globules non-aggregated but they are also distributed, forming a kind of network that enhances physical stability. Sometimes homogenization pressures are also capable of reducing the microbial load of the product prior to the thermal treatment, if they are greater than 200 MPa (N. Cruz et al., 2007; Pereda, Ferragut, Quevedo, Guamis, & Trujillo, 2007; Valencia-Flores et al., 2013). Despite the advantages, this emergent technology is being only used in a laboratory and pilot-scale due to the high investment costs. Hence, as can be seen in Table 2, the vegetable *milks* processing industry normally use hydrocolloids and emulsifiers to prevent phase-separation in developed products.

4 Fermentation of vegetable *milks*

Besides the direct consumption of vegetable *milks*, they might be also used as raw materials to develop yogurt-type products, as has been done with soy *milk*. Hence, these newly fermented products would satisfy a market sector focused on the current consumers' demand of non-dairy products.

In this regards, nut and cereal *milks* are considered as good substrates for the growth of different strains, owing to the presence of non-digestible

components with prebiotic properties in both vegetable matrices. Thus, starch and fiber materials are reported to enhance the physical stability of the fermented vegetable *milk* and to promote the survival of the starters used, not only due to their nutritional contribution but also, since fibers are resistant to gastric juices, they act as protective barriers within the human gastrointestinal tract, (Bosnea et al., 2009; Patel, Pandiella, Wang, & Webb, 2004; Perin, Grill, & Schneider, 2000; Wang, Conway, Brown, & Evans, 1999). Nevertheless, the ability of the starter microorganism to grow in these vegetable raw materials varies largely with the strain. Therefore, studies into bacterial survival are required prior to processing the fermented product.

Most of these innovative fermented products found in the literature have been developed by using probiotic bacteria from Bifidobacteria, Lactobacillus and Streptococcus genera. If probiotic bacteria are used as starter microorganisms, the newly designed product would have an added value, owing to the health benefits that these type of bacteria can exert. Although oat is often used as a raw material, other matrices have also been studied, such as almond, hazelnut or rice. A list of these has been summarized in Table 3.

Table 3: Main characteristics of nut and cereal fermented products found in the literature.

Raw material	Probiotic bacteria used	Growth enhancer (Prebiotic) present and/or added	Reference
Malt, barley and wheat	<i>L. plantarum</i> , <i>L. acidophilus</i>	FOS, β -glucan, starch and other dietary fibers	D Charalampopoulos, Pandiella, and Webb, 2002
Wheat, oat and barley	<i>L. plantarum</i>	FOS, β -glucan, starch and other dietary fibers	Dimitris Charalampopoulos and Pandiella, 2010
Oat	<i>L. plantarum</i> <i>L. reuteri</i> , <i>L. acidophilus</i> , <i>B. bifidum</i>	FOS, β -glucan, starch and other dietary fibers β -glucan and other dietary fibers	Salmoron, Fucinos, Charalampopoulos, and Pandiella, 2009 Angelov, Gotcheva, Kancheva, and Hristozova, 2006 ; Gupta, Cox, and Abu-Ghannam, 2010 Martenson, Oste, and Holst, 2002
Rice-based	<i>L. plantarum</i> , <i>L. paracasei</i> ssp. <i>casei</i> , <i>L. acidophilus</i> <i>B. lactis</i>	Inulin, β -glucan and other dietary fibers Starch	Gokavi, Zhang, Huang, Zhao, and Guo, 2005 Boonyaratankomkit and Wongkhaluang, 2000
Maize	<i>L. paracasei</i> , <i>L. casei</i> , <i>L. rhamnosus</i> , <i>L. acidophilus</i> <i>L. reuteri</i> , <i>L. acidophilus</i> , <i>L. rhamnosus</i>	Inulin, starch, pentosan and other dietary fibers Inulin, starch, pentosan and other dietary fibers	McMaster, Kokott, Reid, and Abratt, 2005 Nyauzi, Jooste, Abu, and Beukes, 2010
Maize and barley	<i>L. rhamnosus</i> , other LAB	Starch, pentosan and other dietary fibers	Holland, Wicklund, and Narrhus, 2004
Emmer wheat	<i>L. paracasei</i> , <i>B. lactis</i>	FOS and other dietary fibers Starch and dietary fiber	Coda, Rizzello, and Gobetti, 2010 Correa, Castro, and Saad, 2008
Cocunut	<i>L. rhamnosus</i>	Starch and other dietary fibers	Blaotta, Di Capua, Coppola, and Aponte, 2012
Chestnut	Kefir grain microorganisms with potentially probiotic effects	Starch and other dietary fibers	Cui, Chen, Wang, and Han, 2013
Walnut	<i>B. lactis</i> , <i>S. thermophilus</i>	Starch, FOS and other dietary fibers	Marti, Martinez, Miralles, and Perez, 2005
Tiger nut, almond or hazelnut	<i>L. Lactobacillus</i>		

B: Bifidobacteria

L: Lactobacillus

S: Streptococcus

4.1 Processing of fermented vegetable *milk*

General industrial processing used to develop nut and cereal fermented vegetable products is based on four main steps: the procedure to obtain vegetable *milk* previously commented on above (Figures 1 and 2), conditioning the *milk* until the optimal starters' growing temperature is reached, the inoculation and incubation procedures (fermentation) and cooling to 4 °C. Nevertheless, depending on the raw material, the type of starters used and the final product features, the whole process may differ.

Some additives are frequently introduced into the vegetable matrix, mainly sugars and prebiotics (as growth enhancers), to promote the viability of bacteria and to reduce the length of the fermentation process (in order to avoid cross-contamination problems). Mono and oligosaccharides, some prebiotics such as inulin, β -glucans and dietary fibers have been the growth enhancers most commonly used by different authors (Akalin, Tokusoglu, Gonc, & Aycan, 2007; Gokavi, Zhang, Huang, Zhao, & Guo, 2005; Ozer, Akin, & Ozer, 2005; Rosburg, Boylston, & White, 2010; Sendra et al., 2008). Potentially, prebiotics are naturally present in both cereals and nuts (i.e. dietary fiber); nevertheless, prebiotic compounds are sometimes added in order to increase the product's health benefits or its technological properties, since the majority are able to increase the viscosity of the *milk*. On the other hand, hydrocolloids, such as carrageenan and xanthan gum, are often added to prevent syneresis and, thus, ensure the physical stability of the product during the stated shelf life. Nonetheless, if the raw material used naturally contains these types of compounds, it might not be necessary to add them during the industrial processing (i.e. β -glucans present in oat *milkz*). Prior to the addition of the starter inoculum, pathogen-free *milks* must be conditioned to reach the fermentation temperature. This is usually around 37 °C, but it depends on the optimal growth temperature of the starter bacteria used. With regards to the fermentation time, much longer is usually needed than during standard cow milk yogurt production, since potential pro-

biotic bacteria (type of bacteria currently chosen to develop these foodstuffs due to the added value on the final product) have more complex nutritional requirements (Severson, 1998), especially when growing in vegetable matrices. The reported fermentation times have been found to be around 16-24 h if no growth enhancers are used in the formulation (Dimitris Charalampopoulos & Pandiella, 2010; D Charalampopoulos, Pandiella, & Webb, 2002; Coda, Rizzello, & Gobetti, 2010; Correa, Castro, & Saad, 2008; Blaiotta, Di Capua, Coppola, & Aponte, 2012; Cui, Chen, Wang, & Han, 2013; Gokavi et al., 2005; Gupta, Cox, & Abu-Ghannam, 2010; Martensson, Oste, & Holst, 2002). The fermentation procedure finishes when the pH value of *milks* reaches 4.2-4.5. Immediately afterwards, the fermented products are sealed, cooled to 4 °C and stored at refrigeration temperatures.

The major challenges affecting these fermented products are related to the sensory quality (appearance and texture) and the resistance of the probiotic microorganisms. Most of these fermented products might have physical stability problems caused by phase separation between components (usually proteins coagulate, forming a non-continuous weak gel, and serum separation occurs at the very beginning of storage time or during storage period). This structure can be observed in Figure 4, where the microstructure is shown of an oat fermented milk with probiotic *L. reuteri* and *S. thermophiles* obtained by using CLSM. As can be observed, the aqueous continuous phase is not completely entrapped in the non-continuous protein-fiber network.

The appearance of these products is often very similar to that observed in a low-fat stirred yogurt. To promote physical stability during the acidification process, hydrocolloids are generally used, as has been commented on above. The most common hydrocolloids used as thickening agents are xanthan gum, modified starches, pectin and cellulose derivatives, among others. Nevertheless, other tools have been used to promote better textural properties. For instance, Martensson, Oste, and Holst (2000) selected strains able to produce exopolysaccharides (EPS) in order to obtain oat-based fermented products. Yoghurts with EPS-producing bacterial strains showed higher viscosity and less phase separa-

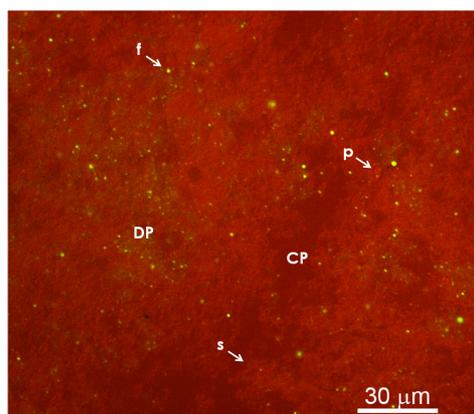


Figure 4: Microstructure obtained by CLSM of fermented oat *milk* with a mixed culture of *L. reuteri* and *S. thermophilus*. (**CP**: Continue phase; **DP**: Dispersed phase; **f**: fat globule; **s**: starters + other fibers). *Source*: personal compilation, University of Valencia

tion in comparison with yoghurts made with strains not producing EPS. This structural property would give rise to a new generation of in situ produced thickeners. This is of general interest, as there is an increasing demand from manufacturers to decrease the addition of stabilizers in yoghurt products.

N. S. Cruz et al. (2009) instead, studied the effect of UHPH treatments (around 250-300 MPa) in soy milk on the fermentation processing. This technology is quite similar to the conventional high-pressure homogenization used in the food industry but considerably higher pressures are applied. Some benefits have been reported for its application in the food industry as it causes interesting changes in structural components and increases the shelf-life of liquid products. The results showed that UHPH soy-yogurts displayed greater firmness and higher water holding capacity than gels produced from conventional homogenized samples.

On the other hand, to enhance the resistance of probiotics during the product's entire shelf-life, new approaches are being taken by different researchers, such as the use of oxygen-impermeable containers, two-step fermentations and the incorporation of micronutrients into the matrices

(peptides and/or amino-acids). Finally, the use of microencapsulation techniques has also been studied as a means of promoting the survival of the probiotic bacteria through the gastrointestinal tract (ability to resist gastric juices and bile) (Soccol et al., 2010).

5 Conclusions

The development of nut and cereal products, unfermented or fermented by means of probiotic bacteria, fully meets the current trend towards an increased consumer demand for healthier products, mainly because of the close relationship between the consumption of vegetable products and the prevention of cancer, atherosclerosis or inflammatory diseases, as has been claimed by some official American and European organizations. In this sense, the demand for and consumption of these products is expected to rise in the next few years, especially in that section of the population which is more aware of health issues. Moreover, and taking into account the low requirements for producing nuts and cereals, the high market potential of these products could be used in the not-to-far future to increase wealth in developing countries by implementing nut and cereals crops such as almonds and oats in non-cultivated lands.

In the development of these products, some important technological deficiencies have been found, mainly related to the product's physical stability during its entire shelf-life. To this end, the optimization of processing techniques must be encouraged and more studies focusing on the microstructure and arrangement of the different components of the products after processing are needed in order to clarify and understand how to improve the appearance and texture of the final product. These studies would also allow a reduction in the amount of additives (hydrocolloids and/or emulsifiers) and thus reduce the economic costs. In the particular case of fermented products, the use of starter microorganisms which are able to both improve the quality (by synthesizing enhanced flavors and providing optimal textures) and exert health benefits for consumers (i.e. probiotics) is the main challenge to be faced in future studies.

Table 4: Main features and health benefits of the raw materials commercially used for producing vegetable *milks*

Raw material	Product features	Health benefits	References
Almond	§ Good source (arranged by importance) of Vit. E, K, Mn, Mg, Cu, P, dietary fiber, riboflavin and protein.	§ Hypocholesterolemic benefits	C.-Y. Chen et al., 2006; Hollis and R. Mattes, 2007; Iacono, Lospalluti, Licastro, and Scalici, 2008; Li et al., 2009; Mandalari, Nueno-Palop, Bisignano, Wickham, and Narbad, 2008; Rajaram, Connell, and Sabate, 2010; Vinson and Cai, 2012
(nut)	§ Good lipid profile, mainly MUFA § Important source of phytonutrients, mainly flavonoids and proanthcyanidins. § Lactose-free; low digestible carbohydrate § Cholesterol-free § High digestibility § Low GI § Low Na content	§ Protective effect against CVD § Antioxidant properties § Electrolite balance contribution § Glucorregulation properties § Prebiotic effect § Anti-inflammatory properties § Possible contribution to protect against some cancers, such as colon cancer § Suitable for celiacs and lactose intolerants	
Chestnut	§ Good source of K, Mg, Fe, Ca, Mn, Cu, dietary fiber, and protein rich in leucine and arginine.	§ Hypocholesterolemic benefits	O. Borges, Goncalves, de Carvalho, Correia, and Silva, 2008; O. P. Borges, Carvalho, Correia, and Silva, 2007; De Vasconcelos, Bennett, Rosa, and Ferreira-Cardoso, 2010
(nut)	§ Low-fat content § Good lipid profile, mainly PUFA and followed by MUFA. § Important amounts folates, thiamin and riboflavin § Important source of phenolic compounds § Lactose-free; low available carbohydrate § Cholesterol-free § High digestibility § Low GI § Low Na content	§ Protective effect against CVD § Antioxidant properties § Electrolite balance contribution § Glucorregulation properties § Prebiotic effect § Possible contribution in protection against some cancers, such as colon cancer § Suitable for celiacs and lactose intolerants	

Table 4 – Continued from previous page...

Raw material	Product features	Health benefits	References
Hazelnut	§ Good source of Vit. E, K, Fe, Ca, Mg, Zn, dietary fiber, and protein rich in arginine and leucine.	§ Hypocholesterolemic benefits	Alasalvar, Shahidi, Liyanapathirana, and Ohshima, 2003; Durak et al., 1999; Koksai, Artik, Simsek, and Gunes, 2006; Li et al., 2009; Mercanligil et al., 2007; Ozdemir et al., 2001, Tey et al., 2011
(nut)	§ Good lipid profile, mainly MUFA. § Important amounts of niacin, Vit.B1, Vit.B2, Vit.B6 and ascorbic acid. § Important source of bioactives and phytochemicals and antioxidant phenolics (mainly caffeic acid) § Lactose-free; low available carbohydrate § Cholesterol-free § High digestibility § Low GI	§ Protective effect against CVD § Antioxidant properties § Electrolite balance contribution § Glucorregulation properties § Prebiotic effect § Possible contribution protection against some cancers, such as colon or prostate cancers § Suitable for celiacs and lactose intolerants	
Walnuts	§ Low Na content § Good source of K, P, Mg, Fe, Mn, Cu, Zn, dietary fiber, and protein rich in arginine.	§ Hypocholesterolemic benefits	Almario, Vonghavaravat, Wong, and Kasim-Karakas, 2001; Banel and F. B. Hu, 2009; Chisholm et al., 1998; Elaine and Feldman, 2002; Li et al., 2009; Sze-Tao and Sathe, 2000; Vinson and Cai, 2012; Zambón et al., 2000
(nut)	§ Good lipid profile, mainly PUFA (linoleic and g-linolenic acids). § Appreciable amounts of Vit.E, niacin, thiamin, riboflavin and folic acid. § Important source of bioactives and phytochemicals § Cholesterol-free § High digestibility § Low GI § Low Na content	§ Protective effect against CVD § Antioxidant properties § Electrolite balance contribution § Glucorregulation properties § Prebiotic effect § Anti-inflammatory properties § Possible contribution in protection against some cancers, such as colon or prostate cancers § Suitable for celiacs and lactose intolerants	

Table 4 – *Continued from previous page...*

Raw material	Product features	Health benefits	References
Amaranth (cereal)	<p>§ Good source of Vit.E, Ca, Mg, K, P, Fe and Zn</p> <p>§ High content in both soluble and insoluble fiber</p> <p>§ Good lipid profile, mainly PUFA (linoleic acid), followed by MUFA (oleic acid)</p> <p>§ Good protein source rich in lysine and methionine</p> <p>§ High phytosterol content, mainly b-sitosterol</p> <p>§ High levels of tocotrienols and squalene (cholesterol-lowering comp.)</p> <p>§ Important amounts of Vit.C, riboflavin and niacin.</p> <p>§ Important source flavonoids and phenolic compounds.</p> <p>§ Source of Lunasin (antitumoral peptide)</p> <p>§ Lactose-free; low available carbohydrate</p> <p>§ Gluten-free</p> <p>§ Cholesterol-free</p> <p>§ High digestibility</p> <p>§ Low GI</p>	<p>§ Hypocholesterolemic benefits</p> <p>§ Protective effect against CVD</p> <p>§ Antioxidant properties</p> <p>§ Electrolite balance contribution</p> <p>§ Glucorregulation properties</p> <p>§ Prebiotic effect</p> <p>§ Possible contribution in protection against some cancers such as colon cancer</p> <p>§ Suitable for celiacs, diabetics and lactose intolerants</p> <p>§ Antitumor effects</p> <p>§ Anti-inflammatory properties</p> <p>§ Anti-anemic effects</p>	<p>Alvarez-Jubete, Arendt, and Gallagher, 2010; Caselato-Sousa and Amaya-Farfan, 2012; Marcone, Kakuda, and Yada, 2003; Sanz-Penella, Wronkowska, Soral-Smietana, and Haros, 2013</p>
Barley (cereal)	<p>§ Good source of Ca, Mg, K, P, Fe and Zn</p> <p>§ High content in dietary fiber, rich in b -glucans</p> <p>§ Good lipid profile, mainly PUFA</p> <p>§ Good protein source rich in lysine</p> <p>§ Important source of tocopherols and tocotrienols (cholesterol-lowering comp.)</p> <p>§ Important amounts of group B vitamins, mainly niacin, riboflavin folate and thiamin.</p>	<p>§ Hypocholesterolemic benefits</p> <p>§ Protective effect against CVD</p> <p>§ Antioxidant properties</p> <p>§ Electrolite balance contribution</p> <p>§ Glucorregulation properties</p> <p>§ Prebiotic effect</p>	<p>AbuMweis, Jew, and Ames, 2010; Ames and Rhymer, 2008; Baik and Ullrich, 2008; K. D. Kaur et al., 2011; Thondre, Ryan, and Henry, 2011; J. L. Ward et al., 2008.</p>

Table 4 – Continued from previous page...

Raw material	Product features	Health benefits	References
Millet (cereal)	<p>§ Good source of Ca, Mg, K, P, Fe Zn</p> <p>§ High content in insoluble dietary fiber</p> <p>§ Good lipid profile, mainly PUFA (linoleic acid), followed by MUFA</p> <p>§ Important amounts of group B vitamins, mainly thiamin, riboflavin, folate and niacin.</p> <p>§ Good source of phenolic compounds</p> <p>§ Lactose-free; low available carbohydrate</p> <p>§ Gluten-free</p> <p>§ Cholesterol-free</p> <p>§ Low GI</p>	<p>§ Hypocholesterolemic benefits</p> <p>§ Protective effect against CVD</p> <p>§ Antioxidant properties</p> <p>§ Glucorregulation properties</p> <p>§ Prebiotic effect</p> <p>§ Possible contribution in protection against some cancers, such as colon cancer</p> <p>§ Suitable for celiacs, diabetics and lactose intolerants</p> <p>§ Anti-anemic effects</p>	<p>Chandrasekara and Shahidi, 2011; Devi, Vijayabharathi, Sathyabama, Malleshi, and Priyadarisini, 2011; Hegde, Rajasekaran, and Chandra, 2005; Léder, 2004; K. D. Kaur et al., 2011;</p>
Oat (cereal)	<p>§ Good source of Vit.E, Ca, Mg, K, P, Fe, Cu and Zn</p> <p>§ High content in dietary fiber, rich in b-glucans</p> <p>§ Good lipid profile, mainly PUFA</p> <p>§ Good protein source rich in sulfur aminoacids, such as methionine and cystine (essential aminoacids)</p> <p>§ Important amounts of group B vitamins, mainly nicacin, riboflavin and thiamin.</p> <p>§ Good source of tocopherols and phenolic compounds</p> <p>§ Lactose-free; low available carbohydrate</p> <p>§ Gluten-free</p>	<p>§ Hypocholesterolemic benefits</p> <p>§ Protective effect against CVD</p> <p>§ Antioxidant properties</p> <p>§ Electrolite balance contribution</p> <p>§ Glucorregulation properties</p> <p>§ Prebiotic effect</p> <p>§ Possible contribution in protection against some cancers, such as colon cancer</p> <p>§ Suitable for celiac, diabetics and lactose intolerants</p>	<p>Biel, Bobko, and Maciorowski, 2009; Daou and Zhang, 2012; Jing and X. Hu, 2012; K. D. Kaur et al., 2011; Kemppainen, Heikkinen, Ristikankare, Kosma, and Julkunen, 2009; Butt, Tahir-Nadeem, Khan, Shabir, and Butt, 2008; Thompson, 2003; J. L. Ward et al., 2008;</p>

Table 4 – Continued from previous page...

Raw material	Product features	Health benefits	References
Quinoa (cereal)	<ul style="list-style-type: none"> § Cholesterol-free § Low Na content § High digestibility § Low GI § Good source of Vit.E, Na, Ca, Mg, K, P, Fe Zn § High content in dietary fiber § Good lipid profile, mainly PUFA (linoleic acid), followed by MUFA(oleic acid) § Good protein source rich in lysine and methionine § High phytosterol content. § High levels of squalene (cholesterol-lowering comp.) § Important amounts of group B vitamins, mainly riboflavin folate and thiamin. § Source of bioactive peptides § Lactose-free; low available carbohydrate § Gluten-free § Cholesterol-free § High digestibility § Low GI 	<ul style="list-style-type: none"> § Antimicrobial and immune effects § Anti-anemic effects § Hypocholesterolemic benefits § Protective effect against CVD § Antioxidant properties § Glucorregulation properties § Prebiotic effect § Possible contribution in protection against some cancers, such as colon cancer § Suitable for celiacs, diabetics and lactose intolerants § Antitumor effects § Anti-inflammatory properties § Anti-anemic effects 	<p>Abugoch James, 2009; Alvarez-Jubete et al., 2010; Brady et al., 2007; Chauhan, Eskin, and Mills, 1999; Zhu et al., 2002.</p>
Rice (cereal)	<ul style="list-style-type: none"> § Good source of K, Ca, Mg, P, Fe, Zn, Se and Si. § Low Na content § Good lipid profile, mainly PUFA (linoleic and g-linolenic acids), followed by MUFA (oleic acid) § Important source of phytosterols, especially b-sitosterol and g-oryzanol (oxidation inhibitors) § Important source of tocopherols (Vit.E) and tocotrienols (LDL-cholesterol lowering) § Important amounts of group B vitamins, mainly nicacin, riboflavin, folate, thiamin, B6 Vit., and panthothenic ac. 	<ul style="list-style-type: none"> § Hypocholesterolemic benefits § Protective effect against CVD § Antioxidant properties § Electrolyte balance contribution § Glucorregulation properties § Suitable for celiacs, and lactose intolerants 	<p>Biswas, Sircar, Mitra, and De, 2011; Cicero and Gaddi, 2001; Facchin, Miotto, do Nascimento Vieira, Barreto, and Amante, 2009; Sierra et al., 2005.</p>

Table 4 – Continued from previous page...

Raw material	Product features	Health benefits	References
Sesame (cereal)	<ul style="list-style-type: none"> § Good source of phenolic compounds § Lactose-free § Gluten-free § Cholesterol-free § High digestibility § Good source of Vit. E Ca, Mg, K, P, Fe, Zn and Cu. 	<ul style="list-style-type: none"> § Antitumor effects § Anti-inflammatory properties. § Hypocholesterolemic benefits 	Cooney, Custer, Okinaka, and Franke, 2001; Sirato-Yasumoto, Katsuta, Okuyama, Takahashi, and Ide, 2001; Wu, Kang, Wang, Jou, and Wang, 2006;
	<ul style="list-style-type: none"> § High content in dietary fiber § Good lipid profile, mainly PUFA (rich in linoleic) § Good protein source § High content in lignan (phytostrogen) § Important amounts of group B vitamins, mainly nicacin, riboflavin folate and thiamin. § Good source of phenolic compounds § Lactose-free; low available carbohydrate § Gluten-free § Cholesterol-free § High digestibility § Low GI 	<ul style="list-style-type: none"> § Protective effect against CVD § Antioxidant properties § Electrolite balance contribution § Glucorregulation properties § Prebiotic effect § Possible contribution in protection against some cancers, such as colon cancer § Suitable for celiacs, diabetics and lactose intolerants 	
Spelt (cereal)	<ul style="list-style-type: none"> § Good source of Vit.E, Ca, Mg, K, P, Fe, Mn and Zn § High content in dietary fiber § Low Na content § Good lipid profile, mainly PUFA (linoleic acid), followed by MUFA (oleic acid) § Good protein source § High levels of, tocopherols and tocotrienols § Good source of group B Vit., mainly thiamin, riboflavin, nicacin, B6 Vit. and folates. § Important source of phenolic compounds. § Lactose-free; low available carbohydrate 	<ul style="list-style-type: none"> § Hypocholesterolemic benefits § Protective effect against CVD § Antioxidant properties § Electrolite balance contribution § Glucorregulation properties § Prebiotic effect § Possible contribution in protection against some cancers, such as colon cancer § Suitable for diabetics and lactose intolerants § Anti-anemic effects 	J. L. Ward et al., 2008; Zielinski, Ceglinska, and Michalska, 2008; Marques et al., 2007, Ruibal-Mendieta et al., 2005; Ranhotra, Gelroth, Glaser, and Lorenz, 1996.

Table 4 – Continued from previous page...

Raw material	Product features	Health benefits	References
Coconut (fruit)	<ul style="list-style-type: none"> § Cholesterol-free § High digestibility § Low GI § Good source of Vit.C and also K, P, Mg and Fe. § Rich in MCFA, mainly lauric acid § Important source of polyphenols § Lactose-free § Cholesterol-free 	<ul style="list-style-type: none"> § Hypocholesterolemic benefits § Protective effect against CVD § Antiviral, antibacterial and antifungal properties § Antioxidant properties § Antithrombotic and antiatherosclerotic effects § Immunostimulatory properties § Suitable for celiacs and lactose intolerants 	DebMandal and Mandal, 2011; Enig, 2004; Raghavendra and Raghavarao, 2010; Sri Lanka Medical Association, 2006.
Tigernut (tuber)	<ul style="list-style-type: none"> § Good source of Vit.E, Ca, Na, K, P, Cu and dietary fiber. § Good lipid profile, mainly MUFA (oleic acid) and followed by linoleic acid. § Important amounts of Vit.C, folates, thiamin and riboflavin § Important source of bioactives and phytochemicals and phenolic compounds. § Lactose-free; low available carbohydrate § Cholesterol-free § High digestibility § Low GI 	<ul style="list-style-type: none"> § Hypocholesterolemic benefits § Protective effect against CVD § Antioxidant properties § Glucoregulation properties § Prebiotic effect § Possible contribution in protection against some cancers, such as colon cancer § Suitable for celiacs and lactose intolerants 	

References

- Abugoch James, L. E. (2009). Chapter 1 quinoa (chenopodium quinoa willd.) *Advances in Food and Nutrition Research*, 1–31. doi:10.1016/s1043-4526(09)58001-1
- AbuMweis, S. S., Jew, S., & Ames, N. P. (2010). Beta-glucan from barley and its lipid-lowering capacity: a meta-analysis of randomized, controlled trials. *European Journal of Clinical Nutrition*, 64(12), 1472–1480. doi:10.1038/ejcn.2010.178
- Adejuyitan, J. (2011). Tigernut processing: its food uses and health benefits. *American Journal of Food Technology*, 6(3), 197–201. doi:10.3923/ajft.2011.197.201
- Agostoni, C., Bresson, J., Tait, S. F., Flynn, A., Golly, I., Korhonen, H., ... Martin, A., et al. (2011). Scientific opinion on the substantiation of health claims related to nuts and essential fatty acids (omega-3/omega-6) in nut oil (id 741, 1129, 1130, 1305, 1407) pursuant to article 13 (1) of regulation (ec) no 1924/2006. *EFSA journal*, 9(4), 2032–1.
- Akalın, A. S., Tokusoglu, O., Gonc, S., & Ayca, S. (2007). Occurrence of conjugated linoleic acid in probiotic yoghurts supplemented with fructooligosaccharide. *International Dairy Journal*, 17(9), 1089–1095. doi:10.1016/j.idairyj.2007.02.005
- Alasalvar, C., Shahidi, F., Liyanapathirana, C., & Ohshima, T. (2003). Turkish tumbul hazelnut (corylus avellana l.). 1. compositional characteristics. *Journal of Agricultural and Food Chemistry*, 51(13), 3790–3796. doi:10.1021/jf0212385
- Almario, R. U., Vonghavaravat, V., Wong, R., & Kasim-Karakas, S. E. (2001). Effects of walnut consumption on plasma fatty acids and lipoproteins in combined hyperlipidemia. *The American Journal of Clinical Nutrition*, 74(1), 72–79. eprint: <http://ajcn.nutrition.org/content/74/1/72.full.pdf+html>. Retrieved from <http://ajcn.nutrition.org/content/74/1/72.abstract>
- Alvarez-Jubete, L., Arendt, E. K., & Gallagher, E. (2010). Nutritive value of pseudocereals and their increasing use as functional gluten-free ingredients. *Trends in Food Science & Technology*, 21(2), 106–113. doi:10.1016/j.tifs.2009.10.014
- Ames, N. P. & Rhymer, C. R. (2008). Issues surrounding health claims for barley. *The Journal of Nutrition*, 138(6), 1237S–1243S. eprint: <http://jn.nutrition.org/content/138/6/1237S.full.pdf+html>
- Angelov, A., Gotcheva, V., Kuncheva, R., & Hristozova, T. (2006). Development of a new oat-based probiotic drink. *International Journal of Food Microbiology*, 112(1), 75–80. doi:<http://dx.doi.org/10.1016/j.ijfoodmicro.2006.05.015>
- Arafat, S., Gaafar, A., Basuny, A., & Nassef, S. (2009). Chufa tubers (cyperus esculentus l.). as a new source of food. *World Applied Sciences Journal*, 7(3), 151–156. Retrieved from [http://www.idosi.org/wasj/wasj7\(2\)/4.pdf](http://www.idosi.org/wasj/wasj7(2)/4.pdf).
- Baik, B.-K. & Ullrich, S. E. (2008). Barley for food: characteristics, improvement, and renewed interest. *Journal of Cereal Science*, 48(2), 233–242. doi:10.1016/j.jcs.2008.02.002
- Banel, D. K. & Hu, F. B. (2009). Effects of walnut consumption on blood lipids and other cardiovascular risk factors: a meta-analysis and systematic review. *American Journal of Clinical Nutrition*, 90(1), 56–63. doi:10.3945/ajcn.2009.27457
- Bernat, N., Chafer, M., A., C., & González-Martínez, C. (2011). Effect of ultra-high pressure homogenization. Effect of ultra-high pressure homogenization and heat treatment on physicochemical properties of almond beverages. In T. P.S., K. V.T., & S. G.D. (Eds.), *Proceedings of the 11th International Congress on Engineering & Food (ICEF11)*. (p. 6). Paper presented at 11th International Congress on Engineering & Food, held at Athens. Athens: ICEF.
- Biel, W., Bobko, K., & Maciorowski, R. (2009). Chemical composition and nutritive value of husked and naked oats grain. *Journal of Cereal Science*, 49(3), 413–418. doi:10.1016/j.jcs.2009.01.009
- Biswas, S., Sircar, D., Mitra, A., & De, B. (2011). Phenolic constituents and antioxidant properties of some varieties of indian

- rice. *Nutrition & Food Science*, 41(2), 123–135. doi:[10.1108/00346651111117391](https://doi.org/10.1108/00346651111117391)
- Blaiotta, G., Di Capua, M., Coppola, R., & Aponte, M. (2012). Production of fermented chestnut purees by lactic acid bacteria. *International Journal of Food Microbiology*, 158(3), 195–202. doi:[10.1016/j.ijfoodmicro.2012.07.012](https://doi.org/10.1016/j.ijfoodmicro.2012.07.012)
- Boonyaratanakornkit, M. & Wongkhalaung, C. (2000). Development of a yoghurt-type product from saccharified rice. *Kasetsart Journal, Natural Sciences*, 34(1), 107–116. Retrieved from http://kasetsartjournal.ku.ac.th/kuj_files/2008/A0804280948236253.pdf
- Borges, O. P., Carvalho, J. S., Correia, P. R., & Silva, A. P. (2007). Lipid and fatty acid profiles of castanea sativa mill. chestnuts of 17 native portuguese cultivars. *Journal of Food Composition and Analysis*, 20(2), 80–89. doi:[10.1016/j.fca.2006.07.008](https://doi.org/10.1016/j.fca.2006.07.008)
- Borges, O., Goncalves, B., de Carvalho, J. L. S., Correia, P., & Silva, A. P. (2008). Nutritional quality of chestnut (castanea sativa mill.) cultivars from portugal. *Food Chemistry*, 106(3), 976–984. doi:[10.1016/j.foodchem.2007.07.011](https://doi.org/10.1016/j.foodchem.2007.07.011)
- Bosnea, L. A., Kourkoutas, Y., Albantaki, N., Tzia, C., Koutinas, A. A., & Kanellaki, M. (2009). Functionality of freeze-dried l. casei cells immobilized on wheat grains. *LWT-food Science And Technology*, 42(10), 1696–1702. doi:[10.1016/j.lwt.2009.05.011](https://doi.org/10.1016/j.lwt.2009.05.011)
- Brady, K., Ho, C.-T., Rosen, R. T., Sang, S., & Karwe, M. V. (2007). Effects of processing on the nutraceutical profile of quinoa. *Food Chemistry*, 100(3), 1209–1216. doi:[10.1016/j.foodchem.2005.12.001](https://doi.org/10.1016/j.foodchem.2005.12.001)
- Burdock, G. A. & Carabin, I. G. (2008). *Breaking down the barriers to functional foods, nutraceuticals and claims*. Elsevier. doi:[10.1016/B978-012373901-8.00007-X](https://doi.org/10.1016/B978-012373901-8.00007-X)
- Butt, M. S., Tahir-Nadeem, M., Khan, M. K. I., Shabir, R., & Butt, M. S. (2008). Oat: unique among the cereals. *European Journal of Clinical Nutrition*, 47(2), 68–79. doi:[10.1007/s00394-008-0698-7](https://doi.org/10.1007/s00394-008-0698-7)
- Canavari, M., Lombardi, P., & Spadoni, R. (2009). Evaluation of the potential interest of italian retail distribution chains for kamut-based products. *Journal of Food Products Marketing*, 16(1), 39–59. Retrieved from <http://ageconsearch.umn.edu/bitstream/7854/1/cp070036.pdf>
- Carlson, J. J., Eisenmann, J. C., Norman, G. J., Ortiz, K. A., & Young, P. C. (2011). Dietary fiber and nutrient density are inversely associated with the metabolic syndrome in us adolescents. *Journal of the American Dietetic Association*, 111(11), 1688–1695. doi:[10.1016/j.jada.2011.08.008](https://doi.org/10.1016/j.jada.2011.08.008)
- Caselato-Sousa, V. M. & Amaya-Farfan, J. (2012). State of knowledge on amaranth grain: a comprehensive review. *Journal of Food Science*, 77(4), R93–R104. doi:[10.1111/j.1750-3841.2012.02645.x](https://doi.org/10.1111/j.1750-3841.2012.02645.x)
- Chan, J. M., Wang, F., & Holly, E. A. (2007). Whole grains and risk of pancreatic cancer in a large population-based case-control study in the san francisco bay area, california. *American Journal of Epidemiology*, 166(10), 1174–1185. doi:[10.1093/aje/kwm194](https://doi.org/10.1093/aje/kwm194)
- Chandrasekara, A. & Shahidi, F. (2011). Bioactivities and antiradical properties of millet grains and hulls. *Journal of Agricultural and Food Chemistry*, 59(17), 9563–9571. doi:[10.1021/jf201849d](https://doi.org/10.1021/jf201849d)
- Charalampopoulos, D. [D], Pandiella, S., & Webb, C. (2002). Growth studies of potentially probiotic lactic acid bacteria in cereal-based substrates. *Journal of Applied Microbiology*, 92(5), 851–859. doi:[10.1046/j.1365-2672.2002.01592.x](https://doi.org/10.1046/j.1365-2672.2002.01592.x)
- Charalampopoulos, D. [Dimitris] & Pandiella, S. S. (2010). Survival of human derived lactobacillus plantarum in fermented cereal extracts during refrigerated storage. *LWT-food Science And Technology*, 43(3), 431–435. doi:[10.1016/j.lwt.2009.09.006](https://doi.org/10.1016/j.lwt.2009.09.006)
- Chauhan, G., Eskin, N., & Mills, P. (1999). Effect of saponin extraction on the nutritional quality of quinoa (chenopodium quinoa willd.) proteins. *Journal of Food Science and Technology*, 36(2), 123–126. Retrieved from <http://cat.inist.fr/>
- Chen, C.-Y., Lapsley, K., & Blumberg, J. (2006). A nutrition and health perspective on almonds. *Journal of the Science of Food*

- and Agriculture, 86(14), 2245–2250. doi:10.1002/jsfa.2659
- Chen, C., Milbury, P., Kwak, H., Collins, F., Samuel, P., & Blumberg, J. (2004). Avenanthramines and phenolic acids from oats are bioavailable and act synergistically with vit c to enhance hamster and human ldl resistance to oxidation. *Journal of Nutrition*, 134(6), 1459–1466. Retrieved from <http://jn.nutrition.org/>
- Chiewchan, N., Phungamngoen, C., & Siriwanayothin, S. (2006). Effect of homogenizing pressure and sterilizing condition on quality of canned high fat coconut milk. *Journal of Food Engineering*, 73(1), 38–44. doi:10.1016/j.foodeng.2005.01.003
- Chisholm, A., Mann, J., Skeaff, M., Frampton, C., Sutherland, W., Duncan, A., & Tiszavari, S. (1998). A diet rich in walnuts favourably influences plasma fatty acid profile in moderately hyperlipidaemic subjects. *European Journal of Clinical Nutrition*, 52(1), 12–16. doi:10.1038/sj.ejcn.1600507
- Cicero, A. & Gaddi, A. (2001). Rice bran oil and gamma-oryzanol in the treatment of hyperlipoproteinaemias and other conditions. *Phytotherapy Research*, 15(4), 277–289. doi:10.1002/ptr.907
- Coda, R., Rizzello, C. G., & Gobbetti, M. (2010). Use of sourdough fermentation and pseudocereals and leguminous flours for the making of a functional bread enriched of gamma-aminobutyric acid (gaba). *International Journal of Food Microbiology*, 137(2-3), 236–245. doi:10.1016/j.ijfoodmicro.2009.12.010
- Coniglio, R. (2008). Frutos secos: el cultivo del almendro. “una actividad alternativa”. *Revista Agromensajes*, 25. Retrieved from <http://www.fcagr.unr.edu.ar/Extension/Agromensajes/25/4AM25.htm>
- Cooney, R., Custer, L., Okinaka, L., & Franke, A. (2001). Effects of dietary sesame seeds on plasma tocopherol levels. *Nutrition and Cancer-an International Journal*, 39(1), 66–71. doi:10.1207/S15327914nc391_9
- Correa, S. B. M., Castro, I. A., & Saad, S. M. I. (2008). Probiotic potential and sensory properties of coconut flan supplemented with lactobacillus paracasei and bifidobacterium lactis. *International Journal of Food Science and Technology*, 43(9), 1560–1568. doi:10.1111/j.1365-2621.2007.01585.x
- CRDO: Consejo Regulador de la Denominación de Origen Chufa de Valencia. (2012). Effects of ultra-high pressure homogenization on microbial and physicochemical shelf life of milk. *Horchata de Chufa, Elaboración*. Retrieved from <http://www.chufadevalencia.org/ver/18/Elaboraci%C3%83%C2%B3n.html>
- Cruz, N. S., Capellas, M., Jaramillo, D. P., Trujillo, A. J., Guamis, B., & Ferragut, V. (2009). Soymilk treated by ultra high-pressure homogenization: acid coagulation properties and characteristics of a soy-yogurt product. *Food Chemistry*, 23(2), 490–496. doi:10.1016/j.foodhyd.2008.03.010
- Cruz, N., Capellas, M., Hernandez, M., Trujillo, A. J., Guamis, B., & Ferragut, V. (2007). Ultra high pressure homogenization of soymilk: microbiological, physicochemical and microstructural characteristics. *Food Research International*, 40(6), 725–732. doi:10.1016/j.foodres.2007.01.003
- Cui, X.-H., Chen, S.-J., Wang, Y., & Han, J.-R. (2013). Fermentation conditions of walnut milk beverage inoculated with kefir grains. *LWT-Food Science and Technology*, 50(1), 349–352. doi:10.1016/j.lwt.2012.07.043
- Daou, C. & Zhang, H. (2012). Oat beta-glucan: its role in health promotion and prevention of diseases. *Comprehensive Reviews in Food Science and Food Safety*, 11(4), 355–365. doi:10.1111/j.1541-4337.2012.00189.x
- De Vasconcelos, M. C. B. M., Bennett, R. N., Rosa, E. A. S., & Ferreira-Cardoso, J. V. (2010). Composition of european chestnut (castanea sativa mill.) and association with health effects: fresh and processed products. *Journal of the Science of Food and Agriculture*, 90(10), 1578–1589. doi:10.1002/jsfa.4016
- de Munter, J. S. L., Hu, F. B., Spiegelman, D., Franz, M., & van Dam, R. M. (2007). Whole grain, bran, and germ intake and risk of type 2 diabetes: a prospective cohort study and systematic review. *Plos*

- medicine*, 4(8), 1385–1395. doi:[10.1371/journal.pmed.0040261](https://doi.org/10.1371/journal.pmed.0040261)
- DebMandal, M. & Mandal, S. (2011). Coconut (*Cocos nucifera* L.: *Arecaceae*): in health promotion and disease prevention. *Asian Pacific Journal of Tropical Medicine*, 4(3), 241–247.
- Del Pozo-Insfran, D., Brenes, C., Saldivar, S., & Talcott, S. (2006). Polyphenolic and antioxidant content of white and blue corn (*zea mays* l.) products. *Food Research International*, 39(6), 696–703. doi:[10.1016/j.foodres.2006.01.014](https://doi.org/10.1016/j.foodres.2006.01.014)
- Devi, P. B., Vijayabharathi, R., Sathyabama, S., Malleshi, N. G., & Priyadarisini, V. B. (2011). Health benefits of finger millet (*eleusine coracana* l.) polyphenols and dietary fiber: a review. *Journal of Food Science and Technology*. doi:[10.1007/s13197-011-0584-9](https://doi.org/10.1007/s13197-011-0584-9)
- Dewanto, V., Wu, X., & Liu, R. (2002). Processed sweet corn has higher antioxidant activity. *Journal of Agricultural and Food Chemistry*, 50(17), 4959–4964. doi:[10.1021/jf0255937](https://doi.org/10.1021/jf0255937)
- Dinelli, G., Segura Carretero, A., Di Silvestro, R., Marotti, I., Fu, S., Benedettelli, S., ... Fernandez Gutierrez, A. (2009). Determination of phenolic compounds in modern and old varieties of durum wheat using liquid chromatography coupled with time-of-flight mass spectrometry. *Journal of Chromatography A*, 1216(43), 7229–7240. doi:[10.1016/j.chroma.2009.08.041](https://doi.org/10.1016/j.chroma.2009.08.041)
- Durak, I., Koksall, I., Kacmaz, M., Buyukkocak, S., Cimen, B., & Ozturk, H. (1999). Hazelnut supplementation enhances plasma antioxidant potential and lowers plasma cholesterol levels. *Clinica Chimica Acta*, 284(1), 113–115. doi:[10.1016/S0009-8981\(99\)00066-2](https://doi.org/10.1016/S0009-8981(99)00066-2)
- Dykes, L. & Rooney, L. W. (2006). Sorghum and millet phenols and antioxidants. *Journal of Cereal Science*, 44(3), 236–251. doi:[10.1016/j.jcs.2006.06.007](https://doi.org/10.1016/j.jcs.2006.06.007)
- Egert, S., Kratz, M., Kannenberg, F., Fobker, M., & Wahrburg, U. (2011). Effects of high-fat and low-fat diets rich in monounsaturated fatty acids on serum lipids, ldl size and indices of lipid peroxidation in healthy non-obese men and women when consumed under controlled conditions. *European Journal of Clinical Nutrition*, 50(1), 71–79. doi:[10.1007/s00394-010-0116-9](https://doi.org/10.1007/s00394-010-0116-9)
- Elaine, B. & Feldman, M. (2002). The scientific evidence for a beneficial health relationship between walnuts and coronary heart disease. *Journal of Nutrition*, 132(5), 1062S–1101S. Retrieved from <http://jn.nutrition.org/>
- Enig, M. G. (2004). Coconut: in support of good health in the 21st century. In *F.A.C.N. Source*. Retrieved from http://coconutoil.com/coconut_oil.21st_century/
- Erra, S. (2012). Process for the preparation of a beverage from nuts. EP Patent 2,342,980 - EP2342980A1. Google Patents. Retrieved from <https://www.google.com/patents/EP2342980B1?cl=en>
- Esmailzadeh, A., Mirmiran, P., & Azizi, F. (2005). Whole-grain consumption and the metabolic syndrome: a favorable association in tehranian adults. *European Journal of Clinical Nutrition*, 59(3), 353–362. doi:[10.1038/sj.ejcn.1602080](https://doi.org/10.1038/sj.ejcn.1602080)
- Faccin, G. L., Miotto, L. A., do Nascimento Vieira, L., Barreto, P. L. M., & Amante, E. R. (2009). Chemical, sensorial and rheological properties of a new organic rice bran beverage. *Rice Science*, 16(3), 226–234. doi:[10.1016/S1672-6308\(08\)60083-9](https://doi.org/10.1016/S1672-6308(08)60083-9)
- Fardet, A. (2010). New hypotheses for the health-protective mechanisms of whole-grain cereals: what is beyond fibre? *Nutrition Research Reviews*, 23(1), 65–134. doi:[10.1017/S0954422410000041](https://doi.org/10.1017/S0954422410000041)
- FDA. U.S. Food and Drug Administration. (2003). Summary of qualified health claims subject to enforcement discretion: nuts and heart disease. Retrieved from <http://www.fda.gov/Food/IngredientsPackagingLabeling/LabelingNutrition/ucm072926.htm>
- Gillingham, L. G., Harris-Janz, S., & Jones, P. J. H. (2011). Dietary monounsaturated fatty acids are protective against metabolic syndrome and cardiovascular disease risk factors. *Lipids*, 46(3), 209–228. doi:[10.1007/s11745-010-3524-y](https://doi.org/10.1007/s11745-010-3524-y)

- Gokavi, S., Zhang, L., Huang, M.-K., Zhao, X., & Guo, M. (2005). Oat-based symbiotic beverage fermented by lactobacillus plantarum, lactobacillus paracasei ssp. casei, and lactobacillus acidophilus. *Journal of Food Science*, 70(4), M216–M223. doi:10.1111/j.1365-2621.2005.tb07191.x
- Granato, D., Branco, G. F., Nazzaro, F., Cruz, A. G., & Faria, J. A. (2010). Functional foods and nondairy probiotic food development: trends, concepts, and products. *Comprehensive Reviews in Food Science and Food Safety*, 9(3), 292–302. doi:10.1111/j.1541-4337.2010.00110.x
- Gupta, S., Cox, S., & Abu-Ghannam, N. (2010). Process optimization for the development of a functional beverage based on lactic acid fermentation of oats. *Biochemical Engineering Journal*, 52(2-3), 199–204. doi:10.1016/j.bej.2010.08.008
- Hegde, P., Rajasekaran, N., & Chandra, T. (2005). Effects of the antioxidant properties of millet species on oxidative stress and glycemic status in alloxan-induced rats. *Nutrition Research*, 25(12), 1109–1120. doi:10.1016/j.nutres.2005.09.020
- Helland, M., Wicklund, T., & Narvhus, J. (2004). Growth and metabolism of selected strains of probiotic bacteria, in maize porridge with added malted barley. *International Journal of Food Microbiology*, 91(3), 305–313. doi:10.1016/j.ijfoodmicro.2003.07.007
- Hollis, J. & Mattes, R. (2007). Effect of chronic consumption of almonds on body weight in healthy humans. *British Journal of Nutrition*, 98(3), 651–656. doi:10.1017/S007114507734608
- Iacono, G., Lospalluti, M., Licastro, G., & Scalici, C. (2008). A new formula based on almond milk for management of cow milk intolerance and food allergies. *Digestive and Liver Disease*, 40(10), A118. Retrieved from [http://www.dldjournalonline.com/article/S1590-8658\(08\)00571-9/fulltext](http://www.dldjournalonline.com/article/S1590-8658(08)00571-9/fulltext)
- Jing, P. & Hu, X. (2012). Nutraceutical properties and health benefits of oats. In *Cereals and pulses* (pp. 21–36). Wiley-Blackwell. doi:10.1002/9781118229415.ch3
- Jones, J. L., Fernandez, M. L., McIntosh, M. S., Najm, W., Calle, M. C., Kalynych, C., ... Lerman, R. H. (2011). A mediterranean-style low-glycemic-load diet improves variables of metabolic syndrome in women, and addition of a phytochemical-rich medical food enhances benefits on lipoprotein metabolism. *Journal of Clinical Lipidology*, 5(3), 188–196. doi:10.1016/j.jacl.2011.03.002
- Kaur, K. D., Jha, A., Sabikhi, L., & Singh, A. (2011). Significance of coarse cereals in health and nutrition: a review. *Journal of Food Science and Technology*, 1–13.
- Kaur, S. & Das, M. (2011). Functional foods: an overview. *Food Science and Biotechnology*, 20(4), 861–875. doi:10.1007/s10068-011-0121-7
- Kelly, J. H., Jr. & Sabate, J. (2006). Nuts and coronary heart disease: an epidemiological perspective. *British Journal of Nutrition*, 96(1, 2), S61–S67. doi:10.1017/BJN20061865
- Kemppainen, T., Heikkinen, M., Ristikankare, M., Kosma, V.-M., & Julkunen, R. (2009). Effect of unkilned and large amounts of oats on nutritional state of celiac patients in remission. *e-SPEN, the European e-Journal of Clinical Nutrition and Metabolism*, 4(1), e30–e34.
- Koksal, A., Artik, N., Simsek, A., & Gunes, N. (2006). Nutrient composition of hazelnut (corylus avellana l.) varieties cultivated in turkey. *Food Chemistry*, 99(3), 509–515. doi:10.1016/j.foodchem.2005.08.013
- Larsson, S., Giovannucci, E., Bergkvist, L., & Wolk, A. (2005). Whole grain consumption and risk of colorectal cancer: a population-based cohort of 60 000 women. *British Journal of Cancer*, 92(9), 1803–1807. doi:10.1038/sj.bjc.6602543
- Léder, I. (2004). Sorghum and millets. *Cultivated Plants, Primarily as Food Sources*. Retrieved from <http://www.eolss.net>
- Li, T. Y., Brennan, A. M., Wedick, N. M., Mantzoros, C., Rifai, N., & Hu, F. B. (2009). Regular consumption of nuts is associated with a lower risk of cardiovascular disease in women with type 2 diabetes. *The Journal of nutrition*, 139(7), 1333–1338.

- Liu, R. H. (2012). Health benefits of phytochemicals in whole foods. *Nutritional Health: Strategies for Disease Prevention*, 293–310.
- Luengo, M. (2009). *La almendra y otros frutos secos: castaña, pistacho, piñon, nuez*. Buenos Aires: Oceano-Ambar.
- Mandalari, G., Nueno-Palop, C., Bisignano, G., Wickham, M. S. J., & Narbad, A. (2008). Potential prebiotic properties of almond (*amygdalus communis* L.) seeds. *Applied and Environmental Microbiology*, 74(14), 4264–4270. doi:10.1128/AEM.00739-08
- Marcone, M. F., Kakuda, Y., & Yada, R. Y. (2003). Amaranth as a rich dietary source of β -sitosterol and other phytosterols. *Plant Foods for Human Nutrition*, 58(3), 207–211.
- Marotti, I., Bregola, V., Aloisio, I., Di Gioia, D., Bosi, S., Di Silvestro, R., ... Dinelli, G. (2012). Prebiotic effect of soluble fibres from modern and old durum-type wheat varieties on lactobacillus and bifidobacterium strains. *Journal of the Science of Food and Agriculture*, 92(10), 2133–2140. doi:10.1002/jsfa.5597
- Marques, C., D'auria, L., Cani, P. D., Baccelli, C., Rozenberg, R., Ruibal-Mendieta, N. L., ... Delzenne, N. M. (2007). Comparison of glycemic index of spelt and wheat bread in human volunteers. *Food Chemistry*, 100(3), 1265–1271. doi:10.1016/j.foodchem.2005.10.003
- Martensson, O., Oste, B., & Holst, O. (2002). The effect of yoghurt culture on the survival of probiotic bacteria in oat-based, non-dairy products. *Food Research International*, 35(8), 775–784. doi:10.1016/S0963-9969(02)00074-1
- Martensson, O., Oste, R., & Holst, O. (2000). Lactic acid bacteria in an oat-based non-dairy milk substitute: fermentation characteristics and exopolysaccharide formation. *Lebensmittel-Wissenschaft Und Technologie-Food Science and Technology*, 33(8), 525–530. doi:10.1006/fstl.2000.0718
- Marti, V., Martinez, O., Miralles, A., & Perez, M. (2005). Product which is fermented without lactose from a shake comprising non-vegetable dried fruits and/or orgeat. WO Patent App. PCT/ES2005/070,053.
- Google Patents. Retrieved from <https://www.google.com/patents/WO2005104862A1?cl=en>
- Marti, V., Martinez, O., Miralles, A., & Perez, M. (2010). Producto fermentado sin lactosa a partir de batido de frutos secos sin legumbres y/o horchata. WO 2005/104862.
- Mattes, R. D., Kris-Etherton, P. M., & Foster, G. D. (2008). Impact of peanuts and tree nuts on body weight and healthy weight loss in adults. *The Journal of nutrition*, 138(9), 1741S–1745S. Retrieved from <http://jn.nutrition.org/>
- McKevith, B. (2004). Nutritional aspects of cereals. *Nutrition Bulletin*, 29(2), 111–142. doi:10.1111/j.1467-3010.2004.00418.x
- McMaster, L., Kokott, S., Reid, S., & Abratt, V. (2005). Use of traditional african fermented beverages as delivery vehicles for bifidobacterium lactis dsm 10140. *International Journal of Food Microbiology*, 102(2), 231–237. doi:10.1016/j.foodmicro.2004.12.013
- Mellen, P. B., Walsh, T. F., & Herrington, D. M. (2008). Whole grain intake and cardiovascular disease: a meta-analysis. *Nutrition Metabolism and Cardiovascular Diseases*, 18(4), 283–290. doi:10.1016/j.numecd.2006.12.008
- Mepba, H. D., Achinewhul, S. C., & Pillay, M. (2009). Stabilised cocosoy beverage: physicochemical and sensory properties. *Journal of the Science of Food and Agriculture*, 86(12), 1839–1846. doi:10.1002/jsfa.2506
- Mercanligil, S. M., Arslan, P., Alasalvar, C., Okut, E., Akgul, E., Pinar, A., ... Shahidi, F. (2007). Effects of hazelnut-enriched diet on plasma cholesterol and lipoprotein profiles in hypercholesterolemic adult men. *European Journal of Clinical Nutrition*, 61(2), 212–220. doi:10.1038/sj.ejcn.1602518
- Murtaugh, M., Jacobs, D., Jacob, B., Steffen, L., & Marquart, L. (2003). Epidemiological support for the protection of whole grains against diabetes. *Proceedings of the Nutrition Society*, 62(1), 143–149. 7th International Vahouny Fibre Symposium, ROYAL COLL PHYS, EDIN-

- BURGH, SCOTLAND, MAY 27-30, 2002. doi:[10.1079/PNS2002223](https://doi.org/10.1079/PNS2002223)
- Myers, J. L. & Allen, J. C. (2012). Nutrition and inflammation insights on dietary pattern, obesity, and asthma. *American Journal of Lifestyle Medicine*, 6(1), 14–17.
- Navarro-Muñoz. (1996). El almendro: variedades y técnicas de cultivo. Andalucía: Consejería de Agricultura y Pesca.
- Nyanzi, R., Jooste, P. J., Abu, J. O., & Beukes, E. M. (2010). Consumer acceptability of a synbiotic version of the maize beverage mageu. *Development Southern Africa*, 27(3), 447–463. doi:[10.1080/0376835X.2010.498955](https://doi.org/10.1080/0376835X.2010.498955)
- Okarter, N. & Liu, R. H. (2010). Health benefits of whole grain phytochemicals. *Critical Reviews in Food Science and Nutrition*, 50(3), 193–208. doi:[10.1080/10408390802248734](https://doi.org/10.1080/10408390802248734)
- Organic Monitor. (2006). *The european market for soya milk & non-dairy drinks*. Dublin: Research & Markets.
- Osca, J. (2007). *Cultivos herbáceos extensivos: cereales*. Valencia: Editorial UPV.
- Ozdemir, M., Ackurt, F., Kaplan, M., Yildiz, M., Loker, M., Gurcan, T., ... Seyhan, F. (2001). Evaluation of new turkish hybrid hazelnut (*corylus avellana* l.) varieties: fatty acid composition, alpha-tocopherol content, mineral composition and stability. *Food Chemistry*, 73(4), 411–415. doi:[10.1016/S0308-8146\(00\)00315-0](https://doi.org/10.1016/S0308-8146(00)00315-0)
- Ozen, A. E., Pons, A., & Tur, J. A. (2012). Worldwide consumption of functional foods: a systematic review. *Nutrition Reviews*, 70(8), 472–481. doi:[10.1111/j.1753-4887.2012.00492.x](https://doi.org/10.1111/j.1753-4887.2012.00492.x)
- Ozer, D., Akin, S., & Ozer, B. (2005). Effect of inulin and lactulose on survival of lactobacillus acidophilus la-5 and bifidobacterium bifidum bb-02 in acidophilus-bifidus yoghurt. *Food Science and Technology International*, 11(1), 19–24. doi:[10.1177/1082013205051275](https://doi.org/10.1177/1082013205051275)
- Packaged Facts. (2012). Dairy alternative beverages in the u.s.: soy milk, almond milk, rice milks and other dairy milk alternatives. Retrieved from <https://www.packagedfacts.com/Soy-Milk-Dairy-6504961/>
- Patel, H., Pandiella, S., Wang, R., & Webb, C. (2004). Influence of malt, wheat, and barley extracts on the bile tolerance of selected strains of lactobacilli. *Food microbiology*, 21(1), 83–89.
- Pereda, J., Ferragut, V., Quevedo, J., Guamis, B., & Trujillo, A. (2007). Effects of ultra-high pressure homogenization on microbial and physicochemical shelf life of milk. *Journal of Dairy Science*, 90(3), 1081–1093. doi:[http://dx.doi.org/10.3168/jds.S0022-0302\(07\)71595-3](http://dx.doi.org/10.3168/jds.S0022-0302(07)71595-3)
- Pereyra, R. & Mutilangi, W. (2012). Method for preparing high acid rtd whole grain beverages. US Patent App. 13/414,184. Google Patents. Retrieved from <http://www.google.com/patents/US20120244249>
- Perrin, S., Grill, J., & Schneider, F. (2000). Effects of fructooligosaccharides and their monomeric components on bile salt resistance in three species of bifidobacteria. *Journal of Applied Microbiology*, 88(6), 968–974. doi:[10.1046/j.1365-2672.2000.01070.x](https://doi.org/10.1046/j.1365-2672.2000.01070.x)
- Phillips, K., Ruggio, D., & Ashraf-Khorassani, M. (2005). Phytosterol composition of nuts and seeds commonly consumed in the united states. *Journal of Agricultural and Food Chemistry*, 53(24), 9436–9445. doi:[10.1021/jf051505h](https://doi.org/10.1021/jf051505h)
- Piergiovanni, A. R., Simeone, R., & Pasqualone, A. (2009). Composition of whole and refined meals of kamut under southern italian conditions. In S. Pierucci (Ed.), *Icheap-9: 9th international conference on chemical and process engineering, pts 1-3* (Vol. 17, pp. 891–896). Chemical Engineering Transactions. 9th International Conference on Chemical and Process Engineering, Rome, ITALY, MAY 10-13, 2009. doi:[10.3303/CET0917149](https://doi.org/10.3303/CET0917149)
- Prior, R. & Gu, L. (2005). Occurrence and biological significance of proanthocyanidins in the american diet. *Phytochemistry*, 66(18), 2264–2280. 4th Tannin Conference held at the Fall Meeting of the American-Chemical-Society, Cellulose & Renewable Mat Div, Philadelphia, PA, 2004. doi:[10.1016/j.phytochem.2005.03.025](https://doi.org/10.1016/j.phytochem.2005.03.025)

- Raghavendra, S. N. & Raghavarao, K. S. M. S. (2010). Effect of different treatments for the destabilization of coconut milk emulsion. *Journal of Food Engineering*, *97*(3), 341–347. doi:[10.1016/j.jfoodeng.2009.10.027](https://doi.org/10.1016/j.jfoodeng.2009.10.027)
- Rajaram, S., Connell, K. M., & Sabate, J. (2010). Effect of almond-enriched high-monounsaturated fat diet on selected markers of inflammation: a randomised, controlled, crossover study. *British Journal of Nutrition*, *103*(6), 907–912. doi:[10.1017/S0007114509992480](https://doi.org/10.1017/S0007114509992480)
- Ranhotra, G., Gelroth, J., Glaser, B., & Lorenz, K. (1996). Nutrient composition of spelt wheat. *Journal of Food Composition and Analysis*, *9*(1), 81–84. doi:<http://dx.doi.org/10.1006/jfca.1996.0009>
- Rosburg, V., Boylston, T., & White, P. (2010). Viability of bifidobacteria strains in yogurt with added oat beta-glucan and corn starch during cold storage. *Journal of Food Science*, *75*(5), C439–C444. doi:[10.1111/j.1750-3841.2010.01620.x](https://doi.org/10.1111/j.1750-3841.2010.01620.x)
- Ruibal-Mendieta, N., Delacroix, D., Mignolet, E., Pycke, J., Marques, C., Rozenberg, R., ... Larondelle, Y. (2005). Spelt (triticum aestivum ssp spelta) as a source of bread-making flours and bran naturally enriched in oleic acid and minerals but not phytic acid. *Journal of Agricultural and Food Chemistry*, *53*(7), 2751–2759. doi:[10.1021/jf048506e](https://doi.org/10.1021/jf048506e)
- Salas-Salvado, J., Casas-Agustench, P., Murphy, M. M., Lopez-Urriarte, P., & Bullo, M. (2008). The effect of nuts on inflammation. *Asia Pacific Journal of Clinical Nutrition*, *17*(1), 333–336. 10th Asian Congress of Nutrition, Taipei, TAIWAN, SEP, 2007. Retrieved from <http://apjcn.nhri.org.tw/>
- Salmeron, I., Fucinos, P., Charalampopoulos, D., & Pandiella, S. S. (2009). Volatile compounds produced by the probiotic strain lactobacillus plantarum ncimb 8826 in cereal-based substrates. *Food Chemistry*, *117*(2), 265–271. doi:[10.1016/j.foodchem.2009.03.112](https://doi.org/10.1016/j.foodchem.2009.03.112)
- Sanchez-Zapata, E., Fernandez-Lopez, J., & Angel Perez-Alvarez, J. (2012). Tiger nut (cyperus esculentus) commercialization: health aspects, composition, properties, and food applications. *Comprehensive Reviews in Food Science and Food Safety*, *11*(4), 366–377. doi:[10.1111/j.1541-4337.2012.00190.x](https://doi.org/10.1111/j.1541-4337.2012.00190.x)
- Sanful, R. E. et al. (2009). Production and sensory evaluation of tigernut beverages. *Pakistan Journal of Nutrition*, *8*(5), 688–690.
- Sanz-Penella, J. M., Wronkowska, M., Soral-Smietana, M., & Haros, M. (2013). Effect of whole amaranth flour on bread properties and nutritive value. *LWT-food Science And Technology*, *50*(2), 679–685. doi:[10.1016/j.lwt.2012.07.031](https://doi.org/10.1016/j.lwt.2012.07.031)
- Saura, F., Cañellas, J., & Soler, L. (1988). La almendra: composición, variedades, desarrollo y maduración. Madrid: Instituto Nacional de Investigaciones Agrarias.
- Scalbert, A. & Williamson, G. (2000). Dietary intake and bioavailability of polyphenols. Retrieved from <http://jn.nutrition.org/>
- Schatzkin, A., Park, Y., Leitzmann, M. F., Hollenbeck, A. R., & Cross, A. J. (2008). Prospective study of dietary fiber, whole grain foods, and small intestinal cancer. *Gastroenterology*, *135*(4), 1163–1167. doi:[10.1053/j.gastro.2008.07.015](https://doi.org/10.1053/j.gastro.2008.07.015)
- Segura, R., Javierre, C., Lizarraga, M. A., & Ros, E. (2006). Other relevant components of nuts: phytosterols, folate and minerals. *British Journal of Nutrition*, *96*(1, 2), S36–S44. doi:[10.1017/BJN20061862](https://doi.org/10.1017/BJN20061862)
- Sendra, E., Fayos, P., Lario, Y., Fernandez-Lopez, J., Sayas-Barbera, E., & Perez-Alvarez, J. A. (2008). Incorporation of citrus fibers in fermented milk containing probiotic bacteria. *Food Microbiology*, *25*(1), 13–21. doi:[10.1016/j.fm.2007.09.003](https://doi.org/10.1016/j.fm.2007.09.003)
- Severson, D. (1998). *Lactic acid fermentations* (E. A. Milwaukee, Ed.). Reed TWNaG (ed). Nutritional requirements of commercially important microorganisms.
- Sharma, A. & Kapoor, A. C. (1996). Levels of antinutritional factors in pearl millet as affected by processing treatments and various types of fermentation. *Plant Food Hum Nutr*, *49*(3), 241–252. doi:[10.1007/bf01093221](https://doi.org/10.1007/bf01093221)
- Sierra, S., Lara-Villoslada, F., Olivares, M., Jimenez, J., Boza, J., & Xaus, J. (2005).

- Increased immune response in mice consuming rice bran oil. *European Journal of Clinical Nutrition*, 44(8), 509–516. doi:10.1007/s00394-005-0554-y
- Sirato-Yasumoto, S., Katsuta, M., Okuyama, Y., Takahashi, Y., & Ide, T. (2001). Effect of sesame seeds rich in sesamin and sesamol on fatty acid oxidation in rat liver. *Journal of Agricultural and Food Chemistry*, 49(5), 2647–2651. doi:10.1021/jf001362t
- Soccol, C. R., Vandenberghe, L. P. d. S., Spier, M. R., Medeiros, A. B. P., Yamaguishi, C. T., Lindner, J. D. D., ... Thomaz-Soccol, V. (2010). The potential of probiotics: a review. *Food Technology and Biotechnology*, 48(4), 413–434. Retrieved from <http://hrcak.srce.hr/file/92463>
- Sri Lanka Medical Association. (2006). Coconuts fats. the ceylon medical journal. Retrieved from <http://www.sljol.info/index.php/CMJ/article/.../1200>
- Stone, D. (2011). Emerging trend of dairy-free almond milk. Food Magazine. Retrieved from <http://www.foodmag.com.au/news/emerging-trend-of-dairy-free-almond-milk>
- Stuckler, D. & Basu, S. (2011). Evaluating the health burden of chronic diseases. In D. Struckler and K. Siegel (Eds.) *Sick societies: Responding to the global challenge of chronic disease*. New York: Oxford University press.
- Sze-Tao, K. & Sathe, S. (2000). Walnuts (*Juglans regia* L): proximate composition, protein solubility, protein amino acid composition and protein in vitro digestibility. *Journal of the Science of Food and Agriculture*, 80(9), 1393–1401. doi:10.1002/1097-0010(200007)80:9<1393::AID-JSFA653>3.0.CO;2-F
- Tey, S. L., Brown, R. C., Chisholm, A. W., Delahunty, C. M., Gray, A. R., & Williams, S. M. (2011). Effects of different forms of hazelnuts on blood lipids and alpha-tocopherol concentrations in mildly hypercholesterolemic individuals. *European Journal of Clinical Nutrition*, 65(1), 117–124. doi:10.1038/ejcn.2010.200
- Thompson, T. (2003). Oats and the gluten-free diet. *Journal of the American Dietetic Association*, 103(3), 376–379. doi:10.1053/jada.2003.50044
- Thondre, P. S., Ryan, L., & Henry, C. J. K. (2011). Barley beta-glucan extracts as rich sources of polyphenols and antioxidants. *Food Chemistry*, 126(1), 72–77. doi:10.1016/j.foodchem.2010.10.074
- Triantafyllou, A. (2002). Non-dairy containing milk substitute products. WO Patent App. PCT/IB2002/001,759. Google Patents. Retrieved from <http://www.google.com/patents/WO2002065855A3?cl=en>
- Valencia-Flores, D. C., Hernandez-Herrero, M., Guamis, B., & Ferragut, V. (2013). Comparing the effects of ultra-high-pressure homogenization and conventional thermal treatments on the microbiological, physical, and chemical quality of almond beverages. *Journal of Food Science*, 78(2), E199–E205. doi:10.1111/1750-3841.12029
- van de Vijver, L. P. L., van den Bosch, L. M. C., van den Brandt, P. A., & Goldbohm, R. A. (2009). Whole-grain consumption, dietary fibre intake and body mass index in the netherlands cohort study. *European Journal of Clinical Nutrition*, 63(1), 31–38. doi:10.1038/sj.ejcn.1602895
- Vinson, J. A. & Cai, Y. (2012). Nuts, especially walnuts, have both antioxidant quantity and efficacy and exhibit significant potential health benefits. *Food & Function*, 3(2), 134–140. doi:10.1039/c2fo10152a
- Wang, X., Conway, P. L., Brown, I. L., & Evans, A. J. (1999). In vitro utilization of amylopectin and high-amylose maize (amylo-maize) starch granules by human colonic bacteria. *Applied and environmental microbiology*, 65(11), 4848–4854. Retrieved from <http://aem.asm.org/>
- Ward, H. A., Keogh, R., Lentjes, M., Luben, R. N., Wareham, N. J., & Khaw, K.-T. (2012). Fibre intake in relation to serum total cholesterol levels and chd risk: a comparison of dietary assessment methods. *European Journal of Clinical Nutrition*, 66(3), 296–304. doi:10.1038/ejcn.2011.184
- Ward, J. L., Poutanen, K., Gebruers, K., Pironen, V., Lampi, A.-M., Nystrom, L., ... Shewry, P. R. (2008). The healthgrain cereal diversity screen: concept, results, and prospects. *Journal of Agricultural and*

- Food Chemistry*, 56(21), 9699–9709. doi:10.1021/jf8009574
- Welch, R. & McConnell, J. (2001). Oats. *Cereals and Cereal Products - Chemistry and Technology*, 367–390. Aspen Publishers Inc., Maryland.
- Whent, M., Huang, H., Xie, Z., Lutterodt, H., Yu, L., Fuerst, E. P., ... Luthria, D. (2012). Phytochemical composition, anti-inflammatory, and antiproliferative activity of whole wheat flour. *Journal of Agricultural and Food Chemistry*, 60(9), 2129–2135. doi:10.1021/jf203807w
- Wu, W.-H., Kang, Y.-P., Wang, N.-H., Jou, H.-J., & Wang, T.-A. (2006). Sesame ingestion affects sex hormones, antioxidant status, and blood lipids in postmenopausal women. *The Journal of nutrition*, 136(5), 1270–1275. Retrieved from <http://jn.nutrition.org/>.
- Zambón, D., Sabaté, J., Muñoz, S., Campero, B., Casals, E., Merlos, M., ... Ros, E. (2000). Substituting walnuts for monounsaturated fat improves the serum lipid profile of hypercholesterolemic men and women: a randomized crossover trial. *Annals of Internal Medicine*, 132(7), 538–546. doi:10.7326/0003-4819-132-7-200004040-00005
- Zhu, N., Sheng, S., Sang, S., Jhoo, J., Bai, N., Karwe, M., ... Ho, C. (2002). Triterpene saponins from debittered quinoa (*Chenopodium quinoa*) seeds. *Journal of Agricultural and Food Chemistry*, 50(4), 865–867. doi:10.1021/jf011002l
- Zielinski, H., Ceglinska, A., & Michalska, A. (2008). Bioactive compounds in spelt bread. *European Food Research and Technology*, 226(3), 537–544. doi:10.1007/s00217-007-0568-1